

Hepatotoxicity and the present herbal hepatoprotective scenario

Priyankar Dey, Manas Ranjan Saha¹, Arnab Sen¹

Department of Zoology, Cellular immunology Laboratory, ¹Department of Botany, Molecular Cytogenetics Laboratory, University of North Bengal, Siliguri, West Bengal, India

Most of the metabolic and physiological processes of our body as well as the detoxification of various drugs and xenobiotic chemicals occur in the liver. During this detoxification process, the reactive chemical intermediates damage the liver severely. There are several commercially available drugs, consumption of which results in idiosyncratic drug reaction mediated hepatotoxicity. Drug induced hepatotoxicity is a burning problem in this regard and several drugs are withdrawn from the market due to their hepatotoxic nature. Today, worldwide search of non-hepatotoxic drugs, especially potent hepatoprotective drugs have led towards the screening of numerous herbal products. Pharmaceutical companies and scientific communities have started to consider the therapeutic efficiency of the plant-based hepatoprotective remedies. Different herbs are mentioned in various ethnopharmacological practices possessing hepatoprotective capacities and around the globe, such herbs are still used by people to cure certain liver diseases. Therefore, we have documented the various aspects of hepatotoxicity and an overview on the current scenario of the hepatoprotective herbal remedies.

Key words: Ayurveda, drug induced hepatotoxicity, ethnopharmacology, hepatoprotective, hepatotoxicity, herbal, idiosyncratic drug reactions, liver

INTRODUCTION

Liver is the largest organ of the body, contributing about 2% of the total body weight in the average adult human. Liver is associated with most of the physiological processes, which include growth, immunity, nutrition, energy metabolism and reproduction. Synthesis and excretion of bile, albumin, prothrombin and production of the compliments, which is the major effector of the humoral branch of the immune system, occurs mainly in the liver. The detoxification of the harmful chemicals occurs in the liver, which in turn, results in various hepatic diseases. Other factors causing liver damage include chronic alcoholism, viral infections, hepatocarcinoma, etc., Drug induced hepatotoxicity (DIHT) resulting in liver damage has turned into a major medical concern in recent years. During DIHT, formation of pro-inflammatory cytokines and reactive free radicals from the hepatic neutrophils and Kupffer cells cause severe oxidative stress.^[1]

Use of herbal remedies as hepatoprotective therapy has been practised in various traditional medicinal systems. A wide variety of ethnomedicinal plants, which have been claimed to be hepatoprotective in the Ayurveda, Siddha, Unani and Amchi medicinal systems as well as in other traditional medicinal practices around the globe, are successfully tested *in vivo* for their therapeutic potential. Therefore, in this review work an effort has been made to highlight the phenomenon of DIHT and the present scenario of herbal drugs and supplements, which have shown promising signs in the amelioration of the hepatic system.

THE HEPATIC SYSTEM

The hepatic system performs hundreds of metabolic and physiological process either individually or in combination with other organs of the body. Lion's share of the protein, carbohydrate and lipid metabolism is performed by the liver. It synthesises various coagulation factors, insulin-like growth factor 1, anti-thrombin, thrombopoietin, angiotensinogen and albumin. Liver is also responsible for the catabolism of bilirubin and various hormones. One of the most important functions of liver is the detoxification of toxic substances. The detoxification process itself very often damages the hepatic system. Though liver possess tremendous regenerative capacity but toxic chemicals and their metabolic intermediates often cause subclinical liver injury.

Access this article online	
Quick Response Code:	Website: www.greenpharmacy.info
	DOI: 10.4103/0973-8258.122046

Address for correspondence: Mr. Priyankar Dey, Department of Zoology, Cellular Immunology Laboratory, University of North Bengal, Siliguri - 734 013, West Bengal, India. E-mail: priyankardey28@gmail.com

Received: 26-08-2013; **Accepted:** 04-09-2013

HEPATOTOXICITY

Liver being closely associated with the gastrointestinal system receives much of the blood from the portal veins, which drains the xenobiotic compounds to the liver. In the liver, the xenobiotic compounds get activated and forms reactive metabolic species (RMS). The RMS through the oxidative stress pathway damage cellular biomolecules, cause protein dysfunctions and damage to the nucleic acids.^[2] Mitochondrial dysfunction results due to RMS mediated disruption of ionic gradients and intracellular Ca²⁺ storage, causing tissue injury. Hepatocellular inflammation is another outcome of DIHT. Activated natural killer T (NKT) cells and Kupffer cells secrete inflammatory mediators such as tumour necrosis factor (TNF)- α , interferon (IFN)- γ and interleukin (IL)-1 β also promotes tissue damage.^[3]

HEPATOTOXICITY CLASSIFICATION

Hepatotoxicity in general can be coined in the following three cases^[4]: (i) the level alanine amino transferase (ALT), that is glutamyl oxalacetic acid transaminase level in the serum increases three-fold, (ii) serum alkaline phosphatase (ALP) level increases two-fold and (iii) serum bilirubin (SBLN) level is also elevated two-fold (when serum ALT and ALP levels also increases).

Hepatotoxicity is of three major classes: (a) Hepatocellular injury: When serum ALT or ALP levels are elevated; (b) Cholestatic injury: When ALP and bilirubin levels in the serum increases; (c) Mixed injury: When both the ALT and ALP levels in the serum increases.^[4]

ROLE OF AYURVEDA IN HEPATOPROTECTION

The Indian Ayurvedic medicinal system dates back to the mid-second millennium BCE describing the usage of various plants based products for the ailment of numerous diseases. About 77 herbal formulations are found in Ayurveda having hepatoprotective properties,^[5] among which many of them have been tested for their hepatoprotective capacity [Table 1]. Many Indian ancient medicinal texts emphasise on the hepatoprotective capacity of certain plants such as Kalmegh, (*Andrographis paniculata*), Bhuia Amla (*Phyllanthus niruri*), Indian bearberry (*Berberis aristata*), Turmeric (*Curcuma longa*), Kutki (*Picrorhiza kurroa*), Mulethi (*Glycyrrhiza glabra*), Punarnava (*Boerhavia diffusa*), Tulsi (*Ocimum sanctum*), Chicory (*Cichorium intybus*), Bhringa Raja (*Eclipta alba*), Kanak champa (*Pterospermum acerifolium*), Guduchi (*Tinispora cordifolia*), Chirayata (*Swertia chirata*), etc.

Table 1: List of some important hepatoprotective medicinal plants mentioned in Ayurveda

Family	Name of plant	Parts used in Ayurveda
Acanthaceae	<i>Andrographis paniculata</i> (Burm.f.) Wall. ex Nees	Whole plant
	<i>Asteracantha longifolia</i> Nees.	Leaf, root and seed
	<i>Hygrophila spinosa</i> T. Ander.	Leaf, root, stem and seed
Asclepiadaceae	<i>Hemidesmus indicus</i> R.Br.	Root
	<i>Gymnema sylvestre</i> (Retz.) R.Br.ex Schult	Leaf
	<i>Cosmpstigma racemosa</i> Roxb.	Root and bark
Asteraceae	<i>Taraxacum officinale</i> F.H. Wigg	Root
	<i>Pyrenthrum indicum</i> DC.	Flower
	<i>Cichorium intybus</i> L.	Whole plant
	<i>Achille millefolium</i> L.	Whole plant
Berberidaceae	<i>Berberis lycium</i> Royle	Leaf
Cucurbitaceae	<i>Bryonia alba</i> Wild Hops	Root
	<i>Luffa echinata</i> Roxb.	Fruit and seed
	Euphorbiaceae	<i>Euphorbia neriifolia</i> L.
<i>Phyllanthus niruri</i> L.		Whole plant
Fumariaceae	<i>Fumaria officinalis</i> L.	Whole plant
	<i>Fumaria parviflora</i> Lamarck.	Whole plant
Guttiferae	<i>Garcinia indica</i> (Linn.) Robs.	Fruit
Gentianaceae	<i>Swertia chirata</i> (Wall.) C. B. Clarke	Whole plant
	<i>Gentiana kurroo</i> Royle	Root
Labiatae	<i>Mentha longifolia</i> (L.) Huds.	Leaf
	<i>Hyssopus officinalis</i> L.	Whole plant
Leguminosae	<i>Trigonella foenumgraecum</i> Linn.	Seed
	<i>Canavalia ensiformis</i> (L.) DC.	Root
Moraceae	<i>Ficus carica</i> L.	Fruit
	<i>Ficus heterophylla</i> L. f.	Root
Meliaceae	<i>Aphanamixis polystachya</i> (Wall.) R.N. Parker	Bark
Myristicaceae	<i>Myristica fragrans</i> Houtt.	Seed
Menispermaceae	<i>Tinispora cordifolia</i> (Willd.) Hook. f.	Stem
	<i>Nelumbo nucifera</i> Gaertn.	Flower
Pinaceae	<i>Pinus roxburghii</i> Sarg.	Oil
Polygonaceae	<i>Rumex crispus</i> L.	Root
Rosaceae	<i>Prunus armeniaca</i> L.	Fruit
Ranunculaceae	<i>Paeonia emodi</i> Wall. ex Royle	Tuber
	<i>Delphinium zaili</i> L.	Whole plant
Rubiaceae	<i>Hedyotis corymbosa</i> (L.) Lam.	Leaf
Solanceae	<i>Lycopersicon esculentum</i> L.	Fruit
Umbelliferae	<i>Apium graveolens</i> L.	Seed
Verbenaceae	<i>Vitex negundo</i> L.	Whole plant
Zingiberaceae	<i>Zingiber officinale</i> Roxb.	Rhizome

At the Rajiv Gandhi Post Graduate Government Ayurvedic Hospital, Paprola, India, a clinical trial was performed, with success, to evaluate the acclaimed hepatoprotective activity of Daruharidra (*Berberis aristata*), Kamachi

(*Solanum nigrum*), Ghrita kumara (*Aloe vera*) and Bhumyamalaki (*Phyllanthus fraternus*).^[6] Presently there are around 89 Ayurvedic formulations used by 37 Indian pharmaceutical companies to prepare hepatoprotective medicines.^[5] The Indian Medicinal Practitioner's Co-operative Pharmacy and Stores have approved some of the drug formulations from the Ayurvedic, Siddha and Unani medicinal system^[7] and they have been listed in Table 2.

PLANTS AS HEPATOPROTECTIVE RESOURCE AND THEIR RECENT PERSPECTIVES

Use of herbal medicine for the treatment of various ailments dated back to thousands of years. Out of the 250,000 higher plant species, about 70,000 species have been used in different traditional medicinal formulations.^[8] Plants have emerged as a great source of pharmaceutical products. In China, the plant-based pharmaceutical industry accounts for one-third of the entire pharmaceutical business and in Malaysia, the annual market of herbal medicine is around 1 billion Malaysian ringgit. In United States alone, the estimated plant-derived pharmaceutical market is of US\$9 billion per year.^[9] There has been increasing scientific and industrial interest in ethnobotanical medicine during the past few decades and thus, the global market of herbal medicine is expected to increase from \$19.5 billion in 2008 to \$32.9 billion in 2013 with an annual growth rate of 11%.^[10] Very often the synthetic drugs and antibiotics are associated with adverse effects, which include hypersensitivity, immunosuppression and allergic reactions. With the emerging cases of hepatotoxicity, antibiotic resistance in bacteria and various side effects, there is a constant need for new and effective therapeutic agents. Thus, the plant-derived bioactive chemicals have drawn the main attention as a source of complementary and alternative medicine. The use of alternative medicines for the treatment of liver diseases has a long history and medicinal plants and their derivatives are extensively used around the

globe for this purpose. Huge interest of the scientific and pharmaceutical community over the therapeutic use of plant-based materials used in various ethnobotanical practices have led to purification and characterisation of various bioactive compounds, which have proven to be hepatoprotective.

Herbal formulations have often been found to work better in a synergistic manner than working alone. One of such formulations is LIV 52, which is a mixture of extracts from *Capparis spinosa*, *Cichorium intybus*, *Solanum nigrum*, *Cassia occidentalis*, *Terminalia arjuna*, *Achillea millefolium*, *Tamarix gallica*, *Eclipta alba*, *Phyllanthus niruri*, *Berberis aristata*, *Taphanus sativus*, *Phyllanthus emblica*, *Plumbago zeylanica*, *Boerhavia diffusa*, *Tinospora cordifolia*, *Embelia ribes*, *Terminalia chebula* and *Fumaria officianlis*.^[11] One of the earliest mentions of the much acclaimed hepatoprotective plant Milk Thistle (*Silybum marianum*) is found in the Bible (Genesis 3:18) when God tells Adam and Eve, 'thorns also and thistles shall it bring forth to thee'. Later on, silymarin, a mixture of isomeric flavolignans-silybin, silydianin and silychristen was isolated from the plant, which has proven to be beneficial in liver related disorders. Similar plants [Table 3] acclaimed in various traditional medicinal practices are now being screened for their hepatoprotective efficiencies.

HEPATOPROTECTIVE PURE ISOLATED COMPOUNDS FROM PLANT SOURCE

Recent trends in the study of hepatoprotective herbal source have turned towards isolation and purification of pure compounds from plants and assessment of their hepatoprotective activity [Table 4]:

Silymarin from *Silybum marianum*, has been a standard hepatoprotective agent for numerous studies and accounts for 180 million US dollars business in Germany alone.^[86] Various bioactive compounds from plant sources possessing antioxidant, anti-cancer, immunostimulatory effect are also being tested for their possible hepatoprotective potential. A wide variety of flavonoids such as quercetin (*Helichrysum arenarium*), myricitoid C (*Cercis siliquastrum*), stachyrin (*Stachys recta*), eupatolin (*Artemisia capillaris*); alkaloids such as atropine (*Datura metel*), pilocarpine (*Aristolochia clementis*), berberine (*Berberis vulgaris*); organic acids and lipids such as glycolic acid (*Cynara scolymus*), dihydrocholic acid (*Curcuma longa*) have shown potent anti-hepatotoxic activity.^[87]

HERBAL HEPATOTOXICITY

Not all herbs are harmless. Many cases of herbal hepatotoxicity have been reported in recent years, which include confirmations of direct hepatic fibrosis, portal

Table 2: A few herbal formulations approved by indian medicinal practitioner's co-operative pharmacy and stores

Ayurvedic	Siddha	Unani
Bhringarajasava	Arumuga chendooram	Jawarish-e-Amilasada
Chandraprabhavati	Annabedhi chendooram 1 and 2	Jawarish-e-Amila luluvi
Drakahadi rasayam	Ayakantha chendooram	Jawarish-e-Tabashir
Guduchi satwam	Mandooradi kudineer	Kurs-e-gul
Jambeeradi panakam	Ayabringaraja karpam	Rue-e-amila
Panchatiktakwatha churnam	Karisalai lehyam	Sherbeth-e-anarshreen
Dhathri loham	Kantha chendooram	Sherbeth-e-deenar
Tapyadi loham	Loha mandooram	Muffarah-e-Ahmedi
Pipilyadi loham	-	Gul-e-Nilofer
Saptamiruda loham	-	Bhoi-Amla

Table 3: A concise list of hepatoprotective plants

Family	Name of plant	Part	Solvent	Important parameters studied	Reference
Acanthaceae	<i>Thunbergia laurifolia</i> Linn.	Leaf	Ethanol	<i>In vitro</i> , <i>in vivo</i> studies; ALT, AST, TGLY, ALP, SBLN, HS, MTT	12
	<i>Hygrophila auriculata</i> Schumach.	Seed	Petroleum ether and methanol	<i>In vivo</i> studies; SGOT, SGPT, ALP, SD, GD, SBLN, HS	13
Asclepiadaceae	<i>Decalepis hamiltonii</i> Wight and Arn.	Root	Water	LP, PC, SOD, CAT, GPX, GR, LG, HS	14
	<i>Tylophora indica</i> (Burm.f.) Merr.	Leaf	Water	<i>in vivo</i> studies; AST, ALT, ALP, TB, LDH, GR, SOD, CAT, GPX, GST, LP, HS	15
Asteraceae	<i>Chamomile capitula</i>	Capitula	Hydro-ethanolic	<i>In vivo</i> studies; LG, NKA, LG, TBARS, ALP, AST, ALT, TP, SBLN, GL	16
	<i>Artemisia absinthium</i> Linn.	Aerial part	Aqueous	<i>In vivo</i> studies; AST, ALT, TNF- α , IL-1, SOD, GPX, MDA, HS	17
	<i>Epaltes divaricata</i> (L.) Cass.	Whole plant	Aqueous	<i>In vivo</i> studies; ALT, AST, ALP, LG, HS	18
Amaranthaceae	<i>Amaranthus spinosus</i> L.	Whole plant	Petroleum ether	<i>In vivo</i> studies; AST, ALT, ALP, SBLN, MDA, GSH, SOD, CAT, HS	19
	<i>Aerva lanata</i> Juss. Ex Schult.	Whole plant	Hydroalcoholic	<i>in vivo</i> studies; AST, ALT, ALP, SBLN	20
Apiaceae	<i>Apium graveolens</i> L.	Seed	Petroleum ether, acetone and methanol	<i>in vivo</i> studies; SGOT, SGPT, ALP, TP, TA	21
Apocynaceae	<i>Nerium indicum</i> Mill.	Flower	Methanolic extract	<i>in vivo</i> studies; AST, ALT, ALP, SBLN, SOD, MDA, HS	22
Asparagaceae	<i>Asparagus racemosus</i> Willd.	Root	Ethanol	<i>in vivo</i> studies; AST, ALT, ALP, SBLN, SOD, CAT, HS	23
	<i>Asparagus racemosus</i> Willd.	Whole plant	Crude extract and aqueous fraction	<i>in vivo</i> ; LP, PO, TBARS	24
Bignoniaceae	<i>Kigelia africana</i> (Lam.) Benth.	Leaves	Aqueous	<i>in vivo</i> studies; ALT, AST, TBARS, SOD, CAT, GPX, DAD	25
Bixaceae	<i>Bixa orellana</i> L.	Whole plant	Methanolic extract	<i>in vivo</i> studies; SGPT, SGOT, CLT, TG, HS	26
Coccolospermaceae	<i>Cochlospermum planchonii</i> Hook.f.	Rhizomes	Aqueous	<i>in vivo</i> studies; SBLN, ALP, ALP	27
Cucurbitaceae	<i>Momordica dioica</i> Roxb. ex Willd.	Leaves	Ethanolic and aqueous	<i>in vivo</i> studies; AST, ALT, ALP, SBLN, HS	28
	<i>Trichosanthes cucumerina</i> L.	Whole plant	Methanol	<i>in vivo</i> studies; ALT, AST, ALP, SBLN, TP, TA, LG, MDA, HS	29
Capparidaceae	<i>Capparis spinosa</i> Linn.	Root	Ethanolic	<i>in vivo</i> studies; ALT, AST, HS	30
Casuarinaceae	<i>Casuarina equisetifolia</i> L.	Whole plant	Methanolic extract	<i>in vivo</i> studies; SGPT, SGOT, CLT, TG, HS	25
Dryopteridaceae	<i>Arachniodes exilis</i> (Hance) Ching	Rhizomes	Ethanol	<i>in vivo</i> studies; SGOT, SGPT, MDA, SOD, LP	31
Euphorbiaceae	<i>Baliospermum montanum</i> Blume	Root	Alcohol and chloroform extract	<i>in vivo</i> studies; SGPT, SGOT, ALP, HS	32
	<i>Croton oblongifolius</i> Roxb.	Aerial part	Petroleum ether, acetone and methanol	<i>in vivo</i> studies; SGOT, SGPT, ALP, TP, TA	33
	<i>Mallotus japonicus</i> (L.f.) Müll. Arg.	Cortex	Aqueous	<i>in vivo</i> ; AST, ALT, SD, GST, GR, GGT, MDA	34
	<i>Phyllanthus niruri</i> L.	Leaves and fruits	Methanol and Aqueous	<i>in vivo</i> ; LP, SGOT, SGPT	35
Fabaceae	<i>Cajanus cajan</i> (L.) Millsp.	Whole plant	Methanolic	<i>in vivo</i> studies; SGPT, SGOT, AST, ALT, CLT, TG, HS	25
	<i>Tephrosia purpurea</i> (Linn.) Pers.	Aerial part	Aqueous-ethanolic extract	<i>in vivo</i> ; AST, ALT, SBLN, ALP, LG, GGT, MDA, HS	36
	<i>Cassia fistula</i> L.	Leaf	N-heptane	<i>in vivo</i> ; SGOT, SGPT, SBLN, ALP	37
	<i>Cassia fistula</i> L.	Seeds	Methanol	<i>in vivo</i> ; SGOT, SGPT, ALP, SBLN	38
	<i>Acacia confusa</i> Merr.	Bark	Hydroalcoholic	<i>in vivo</i> ; AST, ALT, MDA, CP450, SOD, GPX, HS	39
Gentianaceae	<i>Gentiana olivieri</i> Griseb.	Aerial part	Ethanol	<i>in vivo</i> ; MDA, LG, AST, ALT	40
	<i>Halenia elliptica</i> D. Don	Whole plant	Methanol	<i>in vivo</i> ; ALT, AST, ALP, SBLN, HS	41
Juncaceae	<i>Juncus subulatus</i> Forssk.	Tuber	Hydromethanolic	<i>in vivo</i> ; AST, ALT, ALP, TP, TA, TGLY, MDA	42

Contd...

Table 3: Contd...

Family	Name of plant	Part	Solvent	Important parameters studied	Reference
Lamiaceae	<i>Orthosiphon stamineus</i> Benth	Leaves	Methanol extract	<i>in vivo</i> ; AST, ALT, ALP, GST	43
	<i>Hoslundia opposita</i> Vahl	Stem	Methanol and ethyl acetate	<i>in vivo</i> ; AST, ALT, ALP, SBLN	44
	<i>Ocimum sanctum</i> Linn	Leaf	Hydroalcoholic	<i>in vivo</i> ; AST, ALT, ALP, SBLN, HS	45
Liliaceae	<i>Aloe barbadensis</i> Mill.	Aerial parts	Petroleum ether, chloroform and methanol	<i>in vivo</i> ; SGOT, SGPT, ALP, SBLN, TGLY, LP, LG, G6P, and MAH, AND, HS	46
Malvaceae	<i>Hibiscus esculentus</i> Linn.	Root	ethanol	<i>in vivo</i> ; SGPT, SGOT, ALP, SBLN, LP	47
	<i>Hibiscus sabdariffa</i> Linn.	Calyx	Aqueous	<i>in vivo</i> ; ALT, AST, TBARS, SOD, CAT, GPX, DAD	25
Meliaceae	<i>Azadirachta indica</i> A. Juss.	Leaf	Hydromethanolic	<i>in vivo</i> ; GPX, GST, SOD, CAT	48
Myrsinaceae	<i>Embelia ribes</i> Burm.f.	Fruit	Aqueous	<i>in vivo</i> ; SGPT, SGOT, ALP, SBLN, HS	49
Nyctaginaceae	<i>Boerhaavia diffusa</i> Linn.	Root	Aqueous	<i>in vivo</i> ; GOT, GPT, ACP, ALP, GD, SBLN	50
Nelumbonaceae	<i>Nelumbo nucifera</i> Gaertn.	Leaf	Ethanol	<i>in vivo</i> ; ALT, AST, ALP, GGT, SBLN, SOD, CAT, TBARS, LG	51
Piperaceae	<i>Piper chaba</i> Hunter	Fruit	Hydroacetone	<i>in vivo</i> ; AST, ALT, MTT, TNF- α	52
	<i>Piper longum</i> Linn.	Fruits and roots	Milk extract	<i>in vivo</i> ; SGOT, SGPT, ALP, SBLN	53
Pittosporaceae	<i>Pittosporum neelgherrense</i> Wt. and Arn.	Stem bark	Methanolic	<i>in vivo</i> ; AST, ALT, MDA, SOD, GSH, HS	54
Ranunculaceae	<i>Nigella sativa</i> Linn.	Seeds	Aqueous-ethanolic extract	<i>In vitro</i> ; GSH, ALT, AST	55
Rubiaceae	<i>Rubia cordifolia</i> L.	Roots	Aqueous extract	<i>In vivo</i> ; SGOT, SGPT, GGT, ALP, HS	56
Rutaceae	<i>Aegle marmelos</i> (L.) Corr.	fruit pulp/ seeds	Aqueous extract	<i>In vivo</i> ; AST, ALT, ALP, SBLN, HS	57
	<i>Glycosmis pentaphylla</i> Retz.	Whole plant	Methanolic extract	<i>In vivo</i> ; SGPT, SGOT, AST, ALT, CLT, TG, HS	24
	<i>Zanthoxylum armatum</i> DC.	Bark	Ethanol	<i>In vivo</i> ; SGOT, SGPT, ALP, SBLN, TP, SOD, CAT, LG	58
Sterculiaceae	<i>Pterospermum acerifolium</i> Linn.	Leaves	Ethanol	<i>In vivo</i> ; SBLN, TP, SGOT, SGPT, ALP	59
Scrophulariaceae	<i>Scoparia dulcis</i> L.	Whole plant	Methanol, diethyl ether and petroleum ether	<i>In vivo</i> ; AST, ALT, ALP, TP, GLY, LP, SOD, GR, SBLN, HS	60
	<i>Picrorrhiza kurroa</i> Royle ex Benth	Aqueous	Underground stem	<i>In vivo</i> ; AST, ALT, LL, CLT, TGLY	61
Saururaceae	<i>Saururus chinensis</i> (Lour.) Baill.	Whole plant	Ethanol	<i>In vivo</i> ; ALT, AST, MDA, SOD, TC, TGLY, CLT, TA, CE, HS	62
Salvadoraceae	<i>Azima tetraacantha</i> Lam.	Leaves	Ethanol	<i>In vivo</i> ; SGOT, SGPT, SBLN, TP, ALP, TP, TA, CLT, HS	63
Umbelliferae	<i>Bupleurum kanoi</i> Liu	Roots	Ethanol	<i>In vivo</i> and <i>in vitro</i> ; AST, ALT, MDA, SOD, GPX	64
Verbenaceae	<i>Clerodendrum inerme</i> (L.) Gaertn.	Leaves	Ethanol	<i>In vivo</i> ; ALT, AST, ALP, TGLY, CLT	65
	<i>Vitex trifolia</i> L.	Leaves	Ethanol and aqueous	<i>In vivo</i> ; SBLN, TP, ALT, AST, ALP, HS	66
Vitaceae	<i>Vitis vinifera</i> Linn.	Leaves	Ethanol, chloroform, n-butanol, water	<i>In vivo</i> ; MDA, AST LT, GSH, HS	67

SGOT – Serum glutamyl oxalacetic acid transaminase; SGPT – Serum glutamyl pyruvate transaminase; SBLN – Serum bilirubin; GST – Glutathione-S-transferase; HS – Histopathological studies; GPX – Glutathione peroxidase; ALP – Alkaline phosphatase; PC – Protein carbonylation; SD – Sorbitol dehydrogenase; LP – Lipid peroxidation; GD – Glutamate dehydrogenase; SOD – Superoxide Dismutase; CAT – Catalase; AST – Aspartate amino transferase; ALT – Alanine amino transferase; LG – Liver Glutathione; TP – Total Protein; TA – Total albumin; MDA – Malondialdehyde; DAD – d-aminolevulinic acid dehydratase; CLT – Cholesterol; TGLY – Triglycerides; GGT – γ -glutamyl transferase; TBARS – Thiobarbituric reacting substrate; MTT assay – MTT; GR – Glutathione reductase; NKA – Na⁺ K⁺ ATPase activity; GL – Glycogen level; TG – Total glucose; PO – Protein oxidation; MAH – microsomal aniline hydroxylase; G6P – Glucose-6-phosphatase; AND – Amidopyrine N-demethylase; CP450 – Cytochrome P450; TNF- α – Tumor Necrosis factor alpha; LL – Lipoprotein level; CE – Collagen estimation

inflammation, cholestasis, chronic hepatitis, hepatic veno-occlusive and focal hepatic necrosis. Around 60 herbal formulations have been identified as hepatotoxic in 185 recent publications^[88] and the United States Drug-Induced Liver Injury Network have recognised 10% of all DIHT

to be associated with intake of herbal supplements. In countries like India and China, the main problem with herbal hepatotoxicity is that the vast traditional knowledge of both the countries leads people to use herbal supplements without any prior clinical trials, which sometime turns to

Table 4: A concise list of hepatoprotective active compounds

Name of compound	Source	Family	Chemical class	Reference
α -Amyrin and β -Amyrin	<i>Protium heptaphyllum</i>	Burseraceae	Triterpene	68
Anastatin A and Anastatin B	<i>Anastatica hierochuntica</i>	Brassicaceae	Flavonoid	69
Genistein, Orobol and 5,7,4'-trihydroxy-3'-methoxyisoflavone	<i>Erycibe expansa</i>	Leguminosae	Isoflavone	70
γ -Amyrone, γ -Amyrin, 18 β -hydroperoxy-olean. 12-en-3-one and 3-epi- γ -amyrin	<i>Sedum sarmentosum</i>	Crassulaceae	Triterpene	71
Rutin	<i>Artemisia scoparia</i>	Asteraceae	Flavonoid	72
Rubiadin	<i>Rubia cordifolia</i>	Rubiaceae	Anthraquinone	56
Myristin	<i>Myristica fragrans</i>	Myristicaceae	Cetyl ester	73
Naringenin and wighteone	<i>Cudrania cochinchinensis</i>	Moraceae	Flavonoid	74
Kaempferol and salidroside	<i>Rhodiola sachalinensis</i>	Crassulaceae	Phenolic compound	75
Picroliv	<i>Picrorhiza kurroa</i>	Scrophulariaceae	Iridoid glycoside	76
Gentiopicroside and Sweroside	<i>Swertia japonica</i>	Gentianaceae	Iridoid	77
Tetrahydroswertianolin	<i>Swertia japonica</i>	Gentianaceae	Xanthione	77
Mangiferin	<i>Salacia reticulata</i>	Hippocrateaceae	Phenolic compound	78
Torilin and Torilolone	<i>Cnidium monnieri</i>	Apiaceae	Sesquiterpene	79
Acanthoic acid	<i>Acanthopanax koreanum</i>	Araliaceae	Diterpene	80
18 β -glycyrrhetic acid	<i>Glycyrrhiza uralensis</i>	Fabaceae	Glycyrrhetic acid	81
Lithospermate B	<i>Salvia miltorhiza</i>	Lamiaceae	Caffeic acid	82
Corilagin	<i>Terminalia catappa</i>	Combretaceae	Tannin	83
Neoandrographolide	<i>Andrographis paniculata</i>	Acanthaceae	Diterpene	84
Scropolioside-A	<i>Scrophularia koelzii</i>	Scrophulariaceae	Iridoid glycoside	85

be deleterious for the liver. For instance, with an estimated \$400 billion market of 7000 species of Chinese herbs, at least 10 species of them have been reported to be hepatotoxic. A case of severe hepatotoxicity was reported in a 64-year-old female caused by Indian ayurvedic herbal products, Bakuchi (*Psoralea corylifolia*), Khadin (*Acacia catechu*), Bramhi (*Eclipta alba*) and Usheer (*Vetiveria zizanioidis*).^[88] A detail searchable database of numerous hepatotoxic herbal supplements is given by the National Institute of Health NIH^[89] Some frequently reported herbs having hepatotoxic effects are germander (*Teucrium chamaedrys*), valerian (*Valeriana officinalis*), skullcap (*Scutellaria* sp.), comfrey (*Symphytum* sp.), chaparral (*Larrea tridentata*), mistletoe (*Viscum album*), kava kava (*Pipermet hysticum*), Chinese green tea (*Camellia sinensis*), greater celandine (*Chelidonium majus*), pennyroyal (*Menthapulegium*), margosa oil (*Azadirachza indica*), borage (*Borago officinalis*), gotu kola (*Centella asiatica*), broom corn (*Sorghum vulgare*), etc.^[90] Toxic alkaloids from ackee fruit (*Blighia sapida*), *Callilepis laureola*, *Erythroxyton coca*, *Cassia angustifolia*, *Borago officinalis* and *Sassafras albidum* also have been implicated in various hepatotoxic cases. Perhaps, the most inglorious of them all is Kava Kava, which is utilised as sedative throughout the south Pacific Polynesian cultures. A toxic alkaloid pipermethystine found in the stem and the leaves of Kava demonstrated positive signs of hepatotoxicity. Many commercial herbal products such Herbalife®, Enzyte® and Hydroxy cut® have also been reported to render toxic effects on the liver and therefore, the herbal formulations too must undergo proper clinical trials before commercialising.

CONCLUSION

Maintenance of a healthy liver is essential for the overall wellbeing of an individual. Detail clinical diagnostic criteria for assessment of hepatotoxicity has already been published.^[88] The clinical research has confirmed hepatoprotective efficiency of plant-based traditional and alternative medicines and guided the pharmaceutical companies to formulate numerous hepatoprotective drugs. Today, the main problem with the herbal medicines is that many herbs are consumed as polyherbal formulations where multiple herbs work synergistically. The active component responsible for the disease treatment in most cases remains unknown. Therefore, very often some other components of the polyherbal formulations adversely affect the liver. The National Poison Information Service reports that, among 785 cases of adverse reactions by herbal drugs, hepatotoxicity was the most frequent phenomenon.^[91] There are proper guidelines from World Health Organisation (WHO) for the assessment of hepatotoxicity due to herbal products.^[92] A regulated research policy to highlight the advantages of hepatoprotective herbal medicine with respect to their safety and efficacy could result in a better utilisation of these complementary systems of medicine. Thus, we may conclude with the words of Dr S.T. Han,^[93] 'Herbal medicine holds great but still largely unexplored potential for the development of new drugs to combat major health problems' and thus, 'mechanisms for ensuring the safety and control of herbal medicine need to be introduced as part of its formal incorporation into the health service system'.^[8]

ACKNOWLEDGEMENTS

The authors are grateful to Prof. Tapas Kumar Chaudhuri, Cellular Immunology Laboratory, Department of Zoology, University of North Bengal for his constant support and advice during this review work.

REFERENCES

- Liver transplant. Available from: <http://www.nhs.uk/conditions/Liver-transplant/Pages/Introduction.aspx> [Last accessed date 2013 Jan 3].
- Lynch T, Price A. The effect of cytochrome P450 metabolism on drug response, interactions and adverse effects. *Am Fam Physician* 2007;76:391-6.
- Blazka ME, Wilmer JL, Holladay SD, Wilson RE, Luster MI. Role of pro-inflammatory cytokines in acetaminophen hepatotoxicity. *Toxicol Appl Pharmacol* 1995;133:43-52.
- Navarro VJ, Senior JR. Drug-related hepatotoxicity. *N Engl J Med* 2006;354:731-9.
- Kshirsagar AD, Mohite R, Aggrawal AS, Suralkar UR. Hepatoprotective medicinal plants of ayurveda: A review. *Asian J Pharm Clin Res* 2011;4:1-8.
- Sharma YK, Singh H, Mehera BL. Hepatoprotective effect of few herbs in patients receiving antituberculous treatment. *Indian J Tradit Knowl* 2004;3:391-6.
- Thyagarajan SP. Changing therapy: Treatment of viral hepatitis in modern and alternative medicine systems. *IAPJ Pract Pediatr* 1996;4:161-7.
- Guidelines for the Appropriate use of Herbal Medicines: The role of herbal medicines. Available from: <http://apps.who.int/medicinedocs/en/d/Jh2945e/> [Last accessed date 2013 May 12].
- Malaysia's national policy on biological diversity. Available from: <http://www.chm.frim.gov.my/getattachment/9cae4d9c-a772-4858-a899-c21cb2028749/NBP.pdf.aspx?chset=de92b864-ac2b-4e2c-8d0a-afa7076c3663> [Last accessed date 2013 May 16].
- Medicinal plants and extracts. Available from: <http://www.intracen.org/trade-support/medicinal-plants/> [Last accessed date 2013 Jan 27].
- Singh VK, George CX, Gupta KP, Gupta BM. Antiviral activity of plant extract Liv 52 in mice experimentally infected with semliki forest encephalitis virus. *Sci Cult* 1983;49:354-5.
- Pramyothin P, Chirdchupunsare H, Rungsipipat A, Chaichantipyuth C. Hepatoprotective activity of *Thunbergia laurifolia* Linn extract in rats treated with ethanol: *In vitro* and *in vivo* studies. *J Ethnopharmacol* 2005;102:408-11.
- Singh A, Handa SS. Hepatoprotective activity of *Apium graveolens* and *Hygrophila auriculata* against paracetamol and thioacetamide intoxication in rats. *J Ethnopharmacol* 1995;49:119-26.
- Srivastava A, Shivanandappa T. Hepatoprotective effect of the root extract of *Decalepis hamiltonii* against carbon tetrachloride-induced oxidative stress in rats. *Food Chem* 2010;118:411-7.
- Gujrati V, Patel N, Rao VN, Nandakumar K, Gouda TS, Shalam MD. Hepatoprotective activity of alcoholic and aqueous extracts of leaves of *Tylophora Indica* (Linn.) in rats. *Indian J Pharmacol* 2007;39:43-7.
- Gupta AK, Misra N. Hepatoprotective activity of aqueous ethanolic extract of *Chamomile capitula* in paracetamol intoxicated albino rats. *Am J Pharmacol Toxicol* 2006;1:17-20.
- Amat N, Upur H, Blazekovic B. *In vivo* hepatoprotective activity of the aqueous extract of *Artemisia absinthium* L. against chemically and immunologically induced liver injuries in mice. *J Ethnopharmacol* 2010;131:478-84.
- Hewawasam RP, Jayatilaka KA, Pathirana C, Mudduwa LK. Hepatoprotective effect of *Epaltes divaricata* extract on carbon tetrachloride induced hepatotoxicity in mice. *Indian J Med Res* 2004;120:30-4.
- Zeashan H, Amresh G, Singh S, Rao CV. Hepatoprotective activity of *Amaranthus spinosus* in experimental animals. *Food Chem Toxicol* 2008;46:3417-21.
- Manoharan S, Jaswanth A, Sengottuvelu S, Nandhakumar J, Duraisamy R, Karthikeyan D, et al. Hepatoprotective activity of *Aerva Lanata* Linn against paracetamol induced hepatotoxicity in rats. *Res J Pharm Tech* 2008;1:398-400.
- Ahmed B, Alam T, Varshney M, Khan SA. Hepatoprotective activity of two plants belonging to the apiaceae and the euphorbiaceae family. *J Ethnopharmacol* 2002;79:313-6.
- Singhal KG, Gupta GD. Hepatoprotective and antioxidant activity of methanolic extract of flowers of *Nerium oleander* against CCl₄ induced liver injury in rats. *Asian Pac J Trop Med* 2012;5:677-85.
- Rahiman F, Kumar R, Mani T, Niyas M, Kumar S, Phaneendra P, et al. Hepatoprotective activity of *Asparagus racemosus* root on liver damage caused by paracetamol in rats. *Indian J Novel Drug Del* 2011;3:112-7.
- Kamat JP, Boloor KK, Devasagayam TP, Venkatachalam SR. Antioxidant properties of *Asparagus racemosus* against damage induced by G-Radiation in rat liver mitochondria. *J Ethnopharmacol* 2000;71:425-35.
- Olaleye MT, Rocha BT. Acetaminophen-induced liver damage in mice: Effects of some medicinal plants on the oxidative defence system. *Exp Toxicol Pathol* 2008;59:319-27.
- Ahsan MR, Islam KM, Bulbul IJ, Musaddik MA, Haque E. Hepatoprotective activity of methanol extract of some medicinal plants against carbon tetrachloride-induced hepatotoxicity in rats. *Eur J Sci Res* 2009;37:302-10.
- Aliyu R, Okoye ZS, Shier WT. The hepatoprotective cytochrome p-450 enzyme inhibitor isolated from the nigerian medicinal plant *Chochlopermumplanchonii* is a zinc salt. *J Ethnopharmacol* 1995;48:89-97.
- Jain A, Soni M, Deb L, Jain A, Roult AP, Gupta VB, et al. Antioxidant and hepatoprotective activity of ethanolic and aqueous extracts of *Momordica dioica* Roxb. leaves. *J Ethnopharmacol* 2008;115:61-6.
- Sathesh Kumar S, Ravi Kumar B, Krishna Mohan G. Hepatoprotective effect of *Trichosanthes cucumerina* var *cucumerina* L. on carbon tetrachloride induced liver damage in rats. *J Ethnopharmacol* 2009;123:347-50.
- Aghela N, Rashidib I, Mombeinia A. Hepatoprotective activity of *Capparis spinosa* root bark against CCl₄ induced hepatic damage in mice. *Iran J Pharm Res* 2007;6:285-29.
- Zhou D, Ruan J, Cai Y, Xiong Z, Fu W, Wei A. Antioxidant and hepatoprotective activity of ethanol extract of *Arachniodes exilis* (Hance) Ching. *J Ethnopharmacol* 2010;129:232-7.
- Wadekar RR, Supale RS, Tewari KM, Patil KS, Jalalpure SS. Screening of roots of *Baliospermum montanum* for hepatoprotective activity against paracetamol induced liver damage in albino rats. *Int J Green Pharm* 2010;2:220-3.
- Chandan BK, Saxena AK, Shukla S, Sharma N, Gupta DK, Singh K, et al. Hepatoprotective activity of *Woodfordia fruticosa* Kurz flowers against carbon tetrachloride induced hepatotoxicity. *J Ethnopharmacol* 2008;119:218-24.
- Lim H, Kim H, Choi H, Oh S, Choi J. Hepatoprotective effects of bergenin, a major constituent of *Mallotus japonicus*, on carbon tetrachloride-intoxicated rats. *J Ethnopharmacol* 2000;72:469-74.
- Harish R, Shivanandappa T. Antioxidant activity and hepatoprotective potential of *Phyllanthus niruri*. *Food Chem* 2006;95:180-5.

36. Khatri A, Garg A, Agrawal SS. Evaluation of hepatoprotective activity of aerial parts of *Tephrosia purpurea* L. And stem bark of *Tecomella undulate*. J Ethnopharmacol 2009;122:1-5.
37. Bhakta T, Mukherjee PK, Mukherjee K, Banerjee S, Mandal SC, Tapan TK, et al. Evaluation of hepatoprotective activity of *Cassia fistula* leaf extract. J Ethnopharmacol 1999;66:277-82.
38. Chaudhari NB, Chittam KP, Patil VR. Hepatoprotective activity of *Cassia fistula* seeds against paracetamol-induced hepatic injury in rats. Arch Pharm Sci Res 2009;1:218-21.
39. Tung YT, Wu JH, Huang CC, Peng HC, Chen YL, Yang SC, et al. Protective effect of *Acacia confusa* bark extract and its active compound gallic acid against carbon tetrachloride-induced chronic liver injury in rats. Food Chem Toxicol 2009;47:1385-92.
40. Orhana DD, Aslana M, Aktay G, Ergunc E, Yesiladaa E, Erguna F. Evaluation of hepatoprotective effect of *Gentiana olivieri* herbs on subacute administration and isolation of active principle. Life Sci 2003;72:2273-83.
41. Huang B, Ban X, He J, Zeng H, Zhang P, Wang Y. Hepatoprotective and antioxidant effects of the methanolic extract from *Halenia elliptica*. J Ethnopharmacol. 2010;1131:276-81.
42. Abdel-Razik AF, Elshamy AS, Nassar MI, El-Kousy SM, Hamdy H. Chemical constituents and hepatoprotective activity of *Juncus subulatus*. Rev Latinoam Quím 2009;37:70-84.
43. Chin JH, Hussin AH, Ismai S. Anti-hepatotoxicity effect of *Orthosiphon stamineus* Benth against acetaminophen-induced liver injury in rats by enhancing hepatic GST activity. Pharmacogn Res 2009;1:53-8.
44. Akah PA, Odo CL. Hepatoprotective effect of the solvent fractions of the stem of *Hoslundia opposita* Vahl (Lamiaceae) against carbon tetrachloride and paracetamol induced liver damage in rats. Int J Green Pharm 2010;4:54-8.
45. Chattopadhyay RR, Sarkar SK, Ganguly S, Medda C, Basu TK. Hepatoprotective activity of *Ocimum sanctum* leaf extract against paracetamol induced hepatic damage in rats. Indian J Pharmacol 1992;24:163-5.
46. Chandan BK, Saxena AK, Shukla S, Sharma N, Gupta DK, Suri KA, et al. Hepatoprotective potential of *Aloe barbadensis* Mill. against carbon tetrachloride induced hepatotoxicity. J Ethnopharmacol 2007;111:560-6.
47. Sunilson JA, Jayaraj P, Mohan MS, Kumari AA, Varatharajan R. Antioxidant and hepatoprotective effect of the roots of *Hibiscus esculentus* Linn. Int J Green Pharm 2008;2:200-3.
48. Chattopadhyay RR, Bandyopadhyay M. Possible mechanism of hepatoprotective activity of *Azadirachta indica* leaf extract against paracetamol-induced hepatic damage in rats: Part III. Indian J Pharmacol 2005;37:184-5.
49. Tabassum N, Agrawal SS. Hepatoprotective activity of *Embelia ribes* against paracetamol induced acute hepatocellular damage in mice. Exp Med 2003;10:43-4.
50. Rawat KS, Mehrotra S, Tripathi SC, Shome U. Hepatoprotective activity of *Boerhaavia diffusa* L. roots-a popular Indian ethnomedicine. J Ethnopharmacol 1997;56:61-6.
51. Bo H, Xiaoquan B, Jingsheng H, Jing T, Jun T, Youwei W. Hepatoprotective and antioxidant activity of ethanolic extracts of edible lotus (*Nelumbo Nucifera* Gaertn.) leaves. Food Chem 2010;120:873-8.
52. Matsuda H, Ninomiya K, Morikawa T, Yasuda D, Yamaguchi I, Yoshikawa M. Hepatoprotective amide constituents from the fruit of *Piper chaba*: Structural requirements, mode of action, and new amides. Bioorg Med Chem 2009;17:7313-23.
53. Patel JA, Shah US. Hepatoprotective activity of *Piper longum*. Traditional milk extract on carbon tetrachloride induced liver toxicity in wistar rats. Boletín Latinoamericano y del Caribe de Plantas Medicinales y Aromáticas 2009;8:121-9.
54. Shi Y, Sun J, He H, Guo H, Zhang S. Hepatoprotective effects of *Ganoderma lucidum* peptides against D-galactosamine-induced liver injury in Mice. J Ethnopharmacol 2008;117:415-9.
55. Daba MH, Abdel-Rahman MS. Hepatoprotective activity of thymoquinone in isolated rat hepatocytes. Toxicol Lett 1998;95:23-9.
56. Rao GM, Rao CV, Pushpangadan P, Shirwaikar A. Hepatoprotective effects of rubiadin, a major constituent of *Rubia cordifolia* Linn. J Ethnopharmacol 2006;103:484-90.
57. Singh R, Rao HS. Hepatoprotective effect of the pulp/seed of *Aegle marmelos* Correa e x. Roxb. against carbon tetrachloride induced liver damage in rats. Int J Green Pharm 2008;2:232-4.
58. Ranawat LS, Bhatt J, Patel J. Hepatoprotective activity of ethanolic extracts of bark of *Zanthoxylum armatum* DC in CCl₄ induced hepatic damage in rats. J Ethnopharmacol 2010;127:777-80.
59. Kharpate S, Vadnerkar G, Jain D, Jain S. Evaluation of hepatoprotective activity of ethanol extract of *Pterospermum acerifolium* Ster leaves. Indian J Pharm Sci 2007;69:850-2.
60. Praveen TK, Dharmaraj S, Bajaj J, Dhanabal SP, Manimaran S, Nanjan MJ, et al. Hepatoprotective activity of petroleum ether, diethyl ether and methanol extract of *Scoparia dulcis* L. against CCl₄-induced acute liver injury in mice. Indian J Pharmacol 2009;41:110-4.
61. Lee HS, Ahn HC, Ku SK. Hypolipidemic effect of water extracts of *Picrorrhiza rhizoma* in PX-407 induced hyperlipemic ICR mouse model with hepatoprotective effects: A prevention study. J Ethnopharmacol 2006;105:380-6.
62. Wang L, Cheng D, Wang H, Di L, Zhou X, Xu T, et al. The hepatoprotective and antifibrotic effects of *Saururus chinensis* against carbon tetrachloride induced hepatic fibrosis in rats. J Ethnopharmacol 2009;126:487-91.
63. Arthika S, Shanthammal Y, Sheryl Igal N, Elankini P, Pramod Reddy G, Gaidhani SN, et al. Hepatoprotective activity of the ethanolic extract of *Azima tetracantha* against paracetamol-induced hepatotoxicity in wistar albino rats. J Adv Pharm Health Res 2011;1:14-20.
64. Wang BJ, Liu CT, Tseng CY, Wu CP, Yu ZR. Hepatoprotective and antioxidant effects of *Bupleurum kaoi* Liu (Chao Et Chuang) extract and its fractions fractionated using supercritical CO₂ on CCl₄-induced liver damage. Food Chem Toxicol 2004;42:609-17.
65. Gopal N, Sengottuvelu S. Hepatoprotective activity of *Clerodendrum inerme* against CCl₄ induced hepatic injury in rats. Fitoterapia 2008;79:24-6.
66. Manjunatha BK, Vidya SM. Hepatoprotective activity of *Vitex trifolia* against CCl₄ induced hepatic damage. Indian J Pharm Sci 2008;70:241-5.
67. Orhan DD, Orhan N, Ergun E, Ergun F. Hepatoprotective effect of *Vitis vinifera* L. leaves on carbon tetrachloride-induced acute liver damage in rats. J Ethnopharmacol 2007;112:145-51.
68. Oliveira FA, Chaves MH, Almeida FR, Lima RC Jr, Silva RM, Maia JL, et al. Protective effect of α - and β -amyrin, a triterpene mixture from *Protium heptaphyllum* (Aubl.) March. trunk wood resin, against acetaminophen-induced liver injury in mice. J Ethnopharmacol 2005;98:103-8.
69. Yoshikawa M, Xu F, Morikawa T, Ninomiya K, Matsuda H. Anastatins A and B, new skeletal flavonoids with hepatoprotective activities from the desert plant *Anastatica hierochuntica*. Bioorg Med Chem Lett 2003;13:1045-9.
70. Matsuda H, Morikawa T, Xu F, Ninomiya K, Yoshikawa M. New isoflavones and pterocarpane with hepatoprotective activity from the stems of *Erycibe expansa*. Planta Med 2004;70:1201-9.
71. Amin H, Mingshi W, Hong YH, Decheng Z, Lee KH. Hepatoprotective triterpenes from *Sedum sarmentosum*. Phytochem 1998;49:2607-10.
72. Janbaz KH, Saeed SA, Gilani AH. Protective effect of rutin on paracetamol and CCl₄-induced hepatotoxicity in rodents. Fitoterapia 2002;73:557-63.

73. Morita T, Jinno K, Kawagishi H, Arimoto Y, Suganuma H, Inakuma T, et al. Hepatoprotective effect of myristin from nutmeg (*Myristica fragrans*) on lipopolysaccharide/d-galactosamine-induced liver injury. *J Agric Food Chem* 2003;51:1560-5.
74. Lin CC, Lee HY, Chang CH, Namba T, Hattori, M. Evaluation of the liver protective principles from the root of *Cudrania cochinchinensis* var. *gerontogea*. *Phytother Res* 2003;10:13-7.
75. Song EK, Kim JH, Kim JS, Cho H, Nan JX, Soku DH, et al. Hepatoprotective phenolic constituents of *Rhodiola sachalinensis* on tacrine-induced cytotoxicity in Hep G2 cells. *Phytother Res* 2003;17:563-5.
76. Visen PK, Shukla B, Patnaik GK, Kaul S, Kapoor NK, Dhawan BN. Hepatoprotective activity of picroliv, the active principle of *Picrorhiza kurroa*, on rat hepatocytes against paracetamol toxicity. *Drug Dev Res* 1991;22:209-19.
77. Hase K, Li J, Basnet P, Xiong Q, Takamura S, Namba T, et al. Hepatoprotective principles of *Swertia japonica* on D-galactosamine/lipopolysaccharide-induced liver injury in mice. *Chem Pharm Bull* 1997;45:1823-7.
78. Yoshikawa M, Ninomiya K, Shimoda H, Nishida N, Matsuda H. Hepatoprotective and antioxidative properties of *Salacia reticulata*: Preventive effects of phenolic constituents on carbon tetrachloride-induced liver injury in mice. *Biol Pharm Bull* 2003;25:72-6.
79. Oh H, Kim JS, Song EK, Cho H, Kim DH, Park SE, et al. Sesquiterpenes with hepatoprotective activity from *Cnidium monnieri* on tacrine-induced cytotoxicity in Hep G2 cells. *Planta Med* 2002;68:748-9.
80. Park EJ, Zhao YZ, Kim YH, Lee JJ, Sohn DH. Acanthoic acid from *Acanthopanax koreanum* protects against liver injury induced by tert-butyl hydroperoxide or carbon tetrachloride *in vitro* and *in vivo*. *Planta Med* 2004;70:321-7.
81. Shim SB, Kim NJ, Kim DH. β -Glucuronidase inhibitory activity and hepatoprotective effect of 18 β -glycyrrhetic acid from the uralensis. *Planta Med* 2002;66:40-3.
82. Hase K, Kasimu R, Basnet P, Kadota S, Namba T. Preventive effect of lithospermate B from *Salvia miltorhiza* on experimental hepatitis induced by carbon tetrachloride or D-galactosamine/lipopolysaccharide. *Planta Med* 1997;63:22-6.
83. Kinoshita S, Inoue Y, Nakama S, Ichiba T, Aniya Y. Antioxidant and hepatoprotective actions of medicinal herb *Terminalia catappa* L. from Okinawa Island and its tannin corilagin. *Phytotherapy* 2007;14:755-62.
84. Chander R, Srivastava V, Tandon JS, Kapoor NK. Antihepatotoxic activity of diterpenes of *Andrographis paniculata* (Kal-megh) against *Plasmodium berghei* induced hepatic damage in *Mastomys natalensis*. *Int J Pharmacogn* 1995;33:135-8.
85. Garg HS, Bhandari SP, Tripathi SC, Patnaik GK, Puri A, Saxena R, et al. Antihepatotoxic and immunostimulant properties of iridoid glycosides of *Scrophularia koelzii*. *Phytother Res* 1994;8:224-8.
86. Pradhan SC, Girish C. Hepatoprotective herbal drug, silymarin from experimental pharmacology to clinical medicine. *Indian J Med Res* 2006;124:491-504.
87. Valan MF, Britto JD, Venkataraman R. Phytoconstituents with hepatoprotective activity. *Int J Chem Sci* 2010;8:1421-32.
88. Teschke R, Eickhoff A, Wolff A, Frenzel C, Schulze J. Herbal hepatotoxicity and WHO global introspection method. *Ann Hepatol* 2013;12:11-21.
89. LiverTox: Clinical and research information on drug-induced liver injury. Available from: <http://www.livertox.nih.gov/> [Last accessed date 2013 Apr 4].
90. Larson AM, Chopra S, Travis AC. Hepatotoxicity due to herbal medications and dietary supplements. Available from: <http://www.uptodate.com/contents/hepatotoxicity-due-to-herbal-medication-s-and-dietary-supplements> [Last accessed date 2013 Mar 28].
91. Shaw D, Leon C, Koleu S, Murray V. Traditional remedies and food supplements. A five-year toxicological study (1991-1995). *Drug Saf* 1997;17:342-56.
92. Guidelines for the assessment of herbal medicines. Available from: http://whqlibdoc.who.int/hq/1991/WHO_TRM_91.4.pdf [Last accessed date 2013 Jan 24].
93. WHO regional director for the western Pacific region working group on herbal medicines. Available from: <http://apps.who.int/medicinedocs/en/d/Jh2945e/14.html> [Last accessed on 2013 Feb 11].

How to cite this article: Dey P, Saha MR, Sen A. Hepatotoxicity and the present herbal hepatoprotective scenario. *Int J Green Pharm* 2013;7:265-73.

Source of Support: Nil, **Conflict of Interest:** None declared.

"Quick Response Code" link for full text articles

The journal issue has a unique new feature for reaching to the journal's website without typing a single letter. Each article on its first page has a "Quick Response Code". Using any mobile or other hand-held device with camera and GPRS/other internet source, one can reach to the full text of that particular article on the journal's website. Start a QR-code reading software (see list of free applications from <http://tinyurl.com/yzlh2tc>) and point the camera to the QR-code printed in the journal. It will automatically take you to the HTML full text of that article. One can also use a desktop or laptop with web camera for similar functionality. See <http://tinyurl.com/2bw7fn3> or <http://tinyurl.com/3ysr3me> for the free applications.