

# Identification of new antidiabetic agents targeting GLUT4 protein using *in silico* analysis

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## Abstract

**Objective:** Diabetes mellitus is a metabolic syndrome that constitutes a major health problem. It is estimated that 246 million people worldwide have diabetes and that 380 million people will be afflicted with diabetes by 2025. In addition, 3.8 million people die each year from diabetes. Natural products offer an advanced starting point in the search for highly specific and potent modulators of bimolecular function as well as novel drugs. In the present study, GLUT4 protein which plays a significant role in protecting  $\beta$ -cells from damage was selected as potential target. **Materials and Methods:** Three-dimensional structure of GLUT4 protein was built using modeller9v9.19. Modeled structure was validated through structure analysis verification server. With the purpose of identifying, the new potential drugs against GLUT4 protein molecular docking studies of 20 natural compounds were carried out using AutoDock. **Results:** The modeled structure has 87.9% residues in the core region. Results of docking studies clearly showed that good binding interactions of the ligand with both the targets at very low energy level. **Conclusion:** Based on the docking energy value, H-bond interaction the compounds hesperidin, fisetin, eriodictyol, wogonin, and chrysin was selected as the most potent compounds for GLUT4 protein. Hence, this present study suggests the consideration of these compounds for further *in vitro* and *in vivo* studies for its development as antidiabetic drugs.

**Key words:** Diabetics, glut4 protein, homology modeling, molecular docking

## INTRODUCTION

Diabetes is one of serious complex conditions which can change the entire body. The main source of energy is blood glucose which is come from the food we eat. Diabetes is a disease that occurs when blood glucose is too high. The World Health Organization reports there are two types of diabetes, i.e., Type 1 diabetes (T1D) and Type 2 diabetes (T2D). T1D also called as juvenile variety of diabetes it was caused by the absolute deficiency of insulin due to the destruction of insulin-producing pancreatic  $\beta$ -cells. T2D is a multifactorial disease it was caused by resistance of insulin with not only hyper insulin emia and hyperglycemia but also atherosclerosis, hypertension, and an abnormal lipid profile.<sup>[1]</sup> There are 90–95% of peoples were diagnosed with T2D.<sup>[2]</sup> India is one among the six countries with the leading amount of diabetes patients and the every year the number was increased. According to the World Health

Organization report, India had 69.1 million diabetes cases in 2015. By 2035, the number of affected people is expected to increase to 592 million globally. About 80% of adults suffer in diabetes in low- and middle-income countries.<sup>[3]</sup> Many reports stated that diabetes has a great impact on the Indian future economy.<sup>[4]</sup> Hence, there is a great concern which needs to be addressed against diabetics. It also stimulates us to search the new and very effective antidiabetic agents. There are many drugs available in the market with very expensive and lots of side effects. Hence, the current situation in India and other countries increases the demand for identifying a

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**Received:** 13-11-2018

**Revised:** 14-12-2018

**Accepted:** 24-12-2018

cost-effective antidiabetic drug with no major side effects that could help the whole diabetic population. The World Health Organization specialist team on diabetes has listed one of its recommendations that traditional methods of treatment for diabetes should be further investigated.<sup>[5]</sup>

From thousands of year's natural products is one best alternative medicine for many diseases.<sup>[6]</sup> In the past 200 years, there is tremendous research has been developed in the field natural product drug discovery. The plant-based drugs are still used today such as salicylic acid, digitoxin, morphine, quinine, and pilocarpine. Since ancient time, the natural products have played a significant role in the traditional treatment of T2D.<sup>[7]</sup> The first identified diabetes dates back to the Ebers papyrus in Egypt around 1500 B.C.<sup>[8]</sup> Later, in India, the early Ayurvedic texts such as the Sushruta Samhita and the Charaka Samhita, which were written in the 4<sup>th</sup>–5<sup>th</sup> century B.C., described the use of approximately 760 and 500 species of medicinal plants, respectively.<sup>[9]</sup> In the present study, we aimed to discover the new plant-based drug that is used for the treatment of diabetes using computer-aided drug design (CADD) approach.

CADD is one of the greatest methods significantly contributed in the field of drug discovery and development in recent years. Drug discovery and development is a time-consuming and expensive process. On average, it takes 10–15 y and \$500–800 million to introduce a drug into the market.<sup>[10]</sup> In this situation CADD, approaches have been extensively used in the pharmaceutical industry to speed up the process. It helped the scientists to focus on the most promising compounds so it can be minimize the synthetic and biological testing efforts. Molecular docking is one of the computational approaches in CADD used to find out the best matches between a receptor and a ligand. It identify the conformations and orientation of the ligand within a binding site and attempts to place the ligand into the binding site in configurations and conformations appropriate for interacting with the receptor.

## MATERIALS AND METHODS

### Datasets

The structures of 20 compounds were downloaded from Pubchem database. The crystal structure of GLUT 4 protein was not available in protein data bank (PDB) database. Hence, the three-dimensional (3D) structure of GLUT4 was modeled using homology modeling approach.

### Template Search and Sequence Alignment

In the present investigation, the protein sequence of GLUT4 was retrieved from UniProtKB/Swiss-prot database (ID P14672).<sup>[11]</sup> The protein sequence consists of 509 amino acids. To find out the suitable for modeling, the GLUT4

protein search with default parameter was performed against the Brookhaven PDB. High percentage of sequence identity and lower e-value these are two main criteria was used to select the correct template. Based on these criteria, the PDB ID 4PYP was selected as the template to generate the model, as it has the identity score of 65%. Quality of the model depends on the identity between the target and template protein. Hence, the alignment between the target and template was carried out using Clustal Omega.<sup>[12]</sup>

### Molecular Modeling of GLUT4 Protein

Homology modeling is a theoretical method that is used to predict the structure of protein from its sequence with an accuracy that is similar to the best results attained experimentally. Homology modeling of the GLUT4 protein was performed based on the crystal structure of 4PYP. This was used as template to build the 3D structure of GLUT 4 protein. The o template coordinate file was retrieved from the protein databank. Using the sequence alignment between the target and template, the 3D model was build using modeller9v9.19 software.<sup>[13,14]</sup> PYMOL was used to visualize the modeled structure.<sup>[15]</sup> All the modeled structures were ranked based on the internal scoring function (DOPE score), and those with the least internal score was selected as best model and utilized for model validation.<sup>[16-18]</sup>

### Assessment of the Model

Quality of the models was assessed with respect to their energy and stereochemical geometry.<sup>[19]</sup> The stereochemical quality of the modeled protein has been validated by the inspection of Phi/Psi distributions of Ramachandran plot from PROCHECK analysis program using structure analysis verification server (SAVS).<sup>[20]</sup> To assess the reliability of the modeled structure of GLUT4 protein, we calculated the root mean square deviation (RMSD) by superimposing it on the template structure using a Chimera.

### Ligands

The 3D structure of 20 selected compounds such as fisetin (5281614), Morin (5281670), eriodictyol (440735), hesperidin (10621), naringenin (932), apigenin (5280704), baicalein (5281605), chrysin (5281607), luteolin (5280445), tangeritin (68077), wogonin (5281703), isorhamnetin (5281654), kaempferol (5280863), rutin (5280805), quercetin (5280343), genestin (5280961), daidzen (5281708), cyanidin (441674), troxerutin (5486699), and delphinidin (68245) was retrieved as SD file from Pubchem database.

### Protein Preparation

To prepare the structure of homology modeled GLUT4 protein for docking studies, the ligand and the water

molecules were removed. Using the `prepare_receptor4.py` script from MGLTools, charges and nonpolar hydrogen atoms were added.

## Ligand Preparation

The structure of compounds was downloaded from Pubchem database. All structures were energy minimized by the MM2 method and converted to `pdb` extension file which is readable at the AutoDock tools (ADT) interface.

## Protein-ligand Interaction using AutoDock

Using the ADT v1.5.4 and AutoDock v4.2 program, the docking analysis was carried out.<sup>[21]</sup> To run the docking, the searching grid extended for target protein was used to and in the ligand, the polar hydrogen was added. Kollman charges were allocated and atomic solvation parameters were added. Polar hydrogen charges of the Gasteiger-type were assigned and the nonpolar hydrogen was merged with the carbons and the internal degrees of freedom and torsions were set. During the docking process, GLUT4 protein was kept as rigid and ligand allowed to move freely. Using the blind docking option, the search was extended to the whole protein. Affinity maps for all the atom types present, as well as an electrostatic map, were computed with a grid spacing of 0.375 Å. The Lamarckian genetic algorithm was used for docking studies. Based on the binding energy, the results were sorted. Based on the (RMSD values), a cluster analysis was carried out with reference to the starting geometry. It was subsequently performed. The lowest energy conformation of the more populated cluster was considered as the most trustable solution.<sup>[21]</sup>

## RESULTS AND DISCUSSION

### Sequence Alignment between Template and Target

Sequence alignment is a way of arranging the protein sequences used to recognize the area of similarity that may be a consequence of functional, structural, or evolutionary relationship between the sequences. For homology, modeling template selection and sequence alignment between the target and template are one of the main criteria. Hence, to select the template PSI-BLAST was performed. Based on the BLAST results, the PDB id: 4PYP which has 65% of similarity with target protein and has the *e*-value of 0.0 (0.0 indicated good similarity with template) was selected as template for further analysis. Alignment between selected target and template was carried out using Clustal Omega. The alignments which included the residues that were conserved in both the template and query sequences were shown in Figure 1. The identical residues between the query and the template sequences were exposed with the same color.

## Homology Modeling of GLUT4 Protein

Homology modeling gives a clear relationship of homology between the target protein sequence and the protein sequence whose structure has been solved. The 3D structure of the protein provides a valuable insight into its molecular function helps to analyze their interactions with suitable substrates or inhibitors. As a result of homology modeling, totally five models were predicted, among them best model structure was identified based on their dope score [Table 1] which was calculated by `model-single.py` command in `modeler` program. The model TvLDH.B99990003 having minimum dope score was considered as the best model of the GLUT4 protein. The modeled structure was shown in Figure 2.

This model has similar structural features to the template protein. The N-terminal and C-terminal domains are recognized which provides valuable insight into molecular function and also enables the protein-protein interaction to be analyzed.

TARGET	MFSGFQQIGSEEDGEPPQQRVTGTLVAVFSAVLGSLQFQYNIQVINAQVIEQSYNIETH	60
Template	-----NEPSSKILTRMLAVGGAVLGSQFYQNTGQVINAQVIEEFYQTTH	48
	* * * * * : * * * * * : * * * * * : * * * * * : * * * * * : * * * * * : * * * * *	
TARGET	LGRQPEGPPSSIPPGTTLMLVAIFSVGGHSSFLIGTSQILGRKRAMLVNVLAV	120
Template	VHRYGE---SILPTTLMLSLVAIFSVGGHSSFLIGTSQILGRKRAMLVNVLAV	104
	* * * * * : * * * * * : * * * * * : * * * * * : * * * * * : * * * * * : * * * * *	
TARGET	LGGSLNGLANAASYEMILGRPLIGAYSGLTSGLVPRYVGETAPTHLREGALTLMLQAI	180
Template	VSAVLNFGKSLKGFEMILLGRPLIGAYSGLTSGLVPRYVGETAPTHLREGALTLMLQAI	164
	* * * * * : * * * * * : * * * * * : * * * * * : * * * * * : * * * * * : * * * * *	
TARGET	VIGILTAQVGLLESLLGTASLMLLGLLTVLPALLQLVLPFCPEPRYLIIQNLQEP	240
Template	VWGLTAQVGLDSLGNLMLLGLLTVLPALLQLVLPFCPEPRFLINNEENRA	224
	* * * * * : * * * * * : * * * * * : * * * * * : * * * * * : * * * * * : * * * * *	
TARGET	RKSLKRLTGAADVSGVLAELKDKRLEERERPLSLQLLGSRTHRQPLIAAVLQLSQQ	300
Template	KSVLKRLRGTADVTHDQKVEESRQKREKVTILELRSPAYRQPLIAAVLQLSQQ	284
	* * * * * : * * * * * : * * * * * : * * * * * : * * * * * : * * * * * : * * * * *	
TARGET	SGINAVFYSTSFETAGVQPAYIAGAVNTVFTLVSLVERAGRRTHLGLAGH	360
Template	SGINAVFYSTSFETAGVQPAYIAGAVNTVFTLVSLVERAGRRTHLGLAGH	344
	* * * * * : * * * * * : * * * * * : * * * * * : * * * * * : * * * * * : * * * * *	
TARGET	CGCALHTVALLLERVPASYSIVATFGVAFETIGRGIPI#IVAEVFSQSPRAAM	420
Template	AGCALHTIALALLQPLMSYLSIVATFGVAFETIGRGIPI#IVAEVFSQSPRAAI	404
	* * * * * : * * * * * : * * * * * : * * * * * : * * * * * : * * * * * : * * * * *	
TARGET	AVAFSMITSNFIQNGFQVAEANSFVFLFAVLLGFFIIFLVFETGRTGDTQIS	480
Template	AVAFSMITSNFIQNGFQVAEANSFVFLFAVLLGFFIIFLVFETGRTGDTQIS	464
	* * * * * : * * * * * : * * * * * : * * * * * : * * * * * : * * * * * : * * * * *	
TARGET	AAFHRTPSLLQEVKPSLELYLGDEND-----	509
Template	SGFRQGGASQSDKTPPEELFHLGLGDSQVLEH#####	504
	: * * * * * : * * * * * : * * * * * : * * * * * : * * * * * : * * * * * : * * * * *	

Figure 1: Alignment between target and template sequence obtained from Clustal Omega

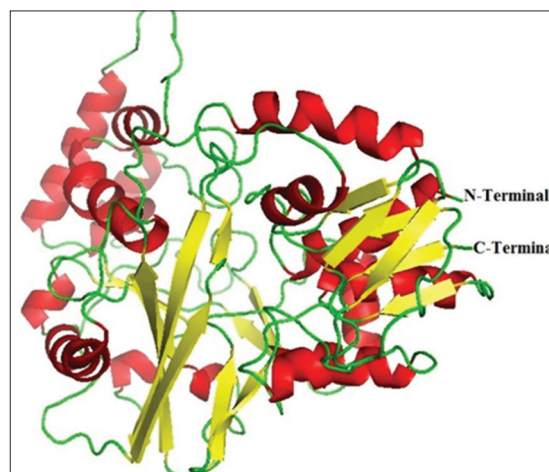


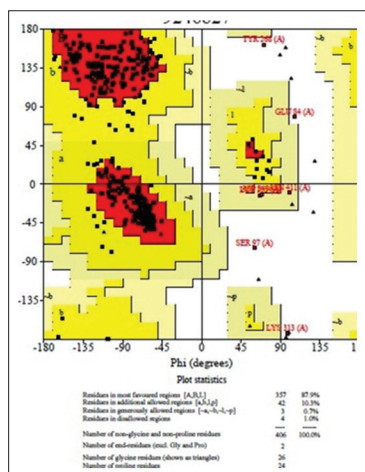
Figure 2: The best-modeled structure of GLUT 4 protein obtained from Modeler 9v9.19. Red color indicates alpha helices, yellow color indicates the beta sheets, and green color indicates the loops

## Model Validation

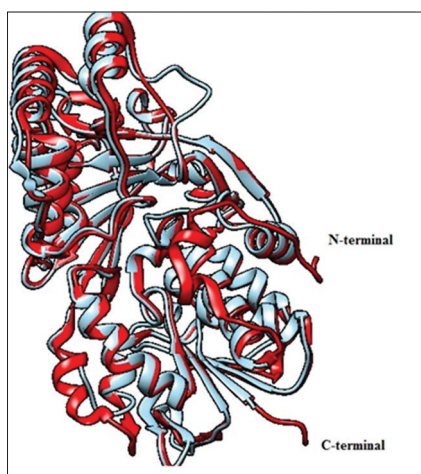
The modeled structures were then subjected to model validation to verify the stereochemical quality of the modeled structure. SAVS was used to validate the modeled structure. A Ramachandran plot calculation was carried out with PROCHECK program. The Phi/Psi distributions of the Ramachandran plot for TvLDH.B99990003.pdb have shown 87.9% of residues in the most favored regions, 10.3% residues in the additionally, 7% residues in generously allowed regions, and 1.0% disallowed regions [Figure 3]. The RMSD value obtained as a result of superimposition using Chimera was 0.097 Å, which indicates that the generated model is quite similar to the template [Figure 4].

## Active Site Prediction

Active site analysis using q site finder module reveals that the compounds were bound in the cavity of the protein containing the following residues VAL 94, TYR 110, PRO 111, VAL 112



**Figure 3:** Ramachandran plot of GLUT4 protein obtained from PROCHECK



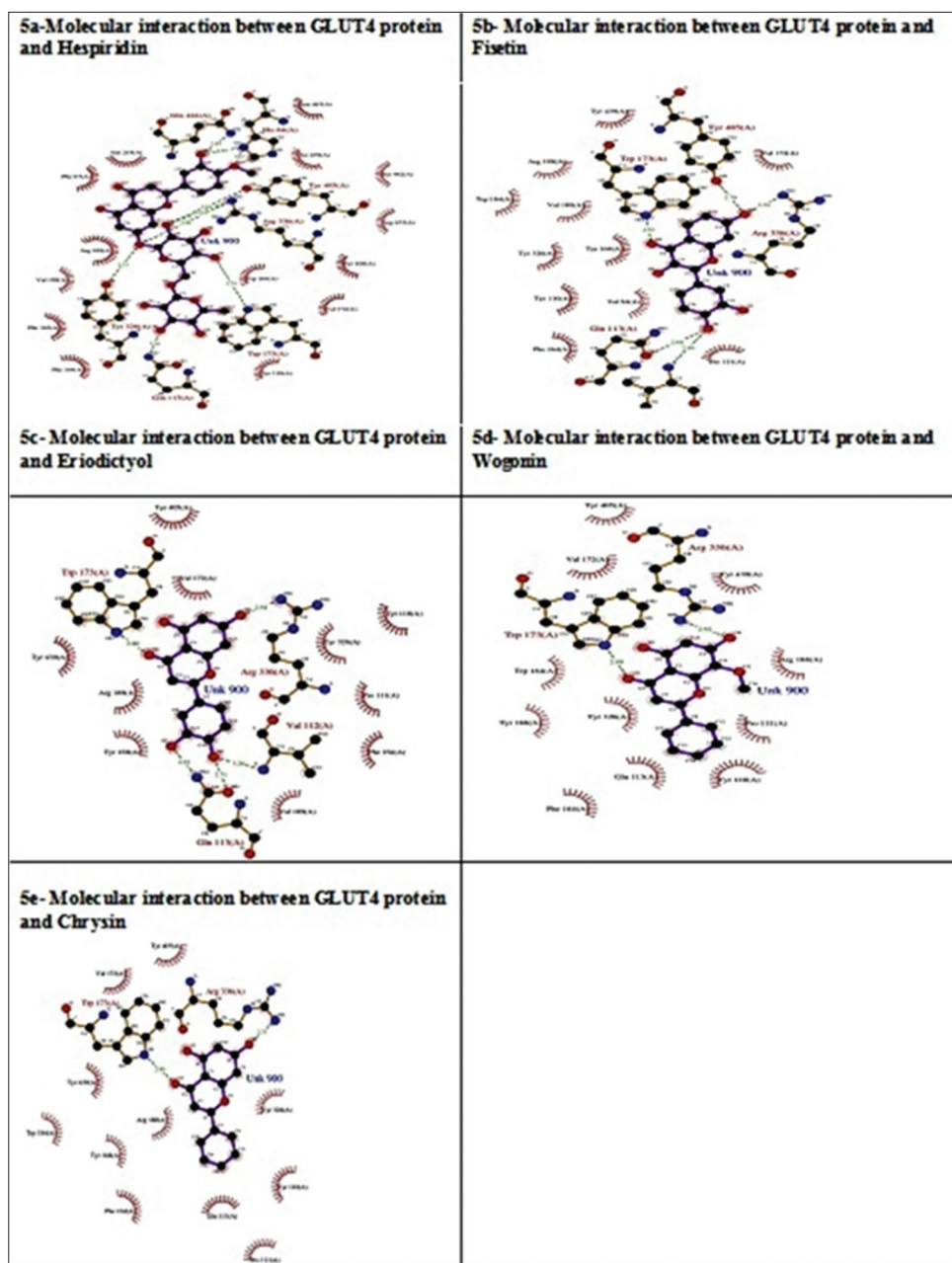
**Figure 4:** Superimposition of modeled GLUT4 protein (target) and template 4PYP

GLN, 113, PHE, 148, PHE 164, PHE 165, TYR 168, THR169, VAL 172, TRP 173, TRP 184, ARG 188, VAL 189, ALA 190, TYR 326, TYR 335, ARG 336, TYR 405, and TYR 439 which were involved in making hydrogen bonds with GLUT4 protein with inhibitors. These amino acids residues play a major role in the binding affinity with the ligand molecule.

## Molecular Docking

To further validate the further quality of the modeled structure and understanding the binding mechanism in terms of affinity as well as selectivity, in this study, molecular docking was carried out. In the predicted active sites, the molecular docking was carried out by the AutoDock program. Results of this docking study showed that selected compounds effectively bind to active site region of GLUT4 protein and it also provides significant results by the least values of the binding energy. This also helped to identify important residues involved in GLUT4-compounds interaction. Results of docking studies showed that there are many conserved amino acid residues in GLUT4 play important role in maintaining a functional protein conformation and also directly involved in binding to GLUT4 protein. As is well known, hydrogen bonds play an essential role in the structure and function of biological molecules. In order gets the H bonding details, LigPlot analysis was carried out. It is used to expose the interaction mechanism between the GLUT4 and selected compounds. The best possible binding affinities and H-bond interaction details of the best five compounds at targeted protein active sites were displayed in Figure 5, and their corresponding energy values were listed in Table 2, respectively.

Analysis of results of the docking studies showed that most of the compounds strongly interact with active site of GLUT4 protein through H-bond interaction. The amino acids, namely HIS-64, VAL-112, GLN-113, TRP-173, TYR-326, ARG-336, TYR-405, and GLN-444 involved in H-bond interaction with the compound. The H-bond involvement used the complex to achieve the established conformation of the complex structure. Hydrogen bond interaction could be acted either antagonist or agonist for a ligand with receptor. In our study, hydrogen bond interaction worked as antagonist. We also noticed that most of the compounds have >3 H-bond interaction with GLUT4 protein, it stated that more number of H-bond showed that the high affinity of ligand with receptor [Table 2]. From these docking studies, we can also able to find out the atoms involved in accepting and donating H-bond. A hydrogen bond donor defined nitrogen or oxygen atom with hydrogen attached; while an acceptor defined nitrogen, oxygen, or fluorine atom with at least one vacant valence to accept a hydrogen atom. In the present study, oxygen with hydrogen was mostly involved in donating H-bond, and nitrogen atom was generally involved in accepting H-bond. From these docking studies, we find out that HIS-64, VAL-112, GLN-113, TRP-173, TYR-326,



**Figure 5:** Molecular interactions of GLUT4 protein and top ranked ligands through H-bond interactions

**Table 1:** Dope score values of successfully produced models

File name	DOPE score
TvLDH.B99990001.pdb	-38013.863216
TvLDH.B99990002.pdb	-37802.813400
TvLDH.B99990003.pdb	-39072.812347
TvLDH.B99990004.pdb	-36487.41724
TvLDH.B99990005.pdb	-36851.152344

ARG-336, TYR-405, and GLN-444 these residues may be functionally important residues used to understand the function of protein. Of 20 compounds hesperidin, fisetin,

eriodictyol, wogonin, and chrysin had the very good binding with GLUT4. Hence, these compounds may be act as lead to identify the new antidiabetic agents.

## CONCLUSION

Of 20 compounds the hesperidin, fisetin, eriodictyol, wogonin, and chrysin were selected as best compounds based on the docking results. Further investigation of the function of the compound will help a better understanding the development of these compounds as an antidiabetic agent.

**Table 2:** List of selected potent compounds and their docking energy value and interactions with the GLUT4 protein

Compound name	Amino acids involved in interaction	Ligand atom involved in H-bond interaction	H-bond distance	Energy value (kcal/mol)	Inhibition constant (nM)
Hesperidin	HIS - 64	ND1	2.87	-8.39	7.83
	GLN - 113	NE2	3.16		
	TRP - 173	NE1	2.72		
	TYR - 326	OH	2.73		
	ARG - 336	NH1	3.08		
	TYR - 405	OH	2.84		
	GLN - 444	NE2	2.85		
Fisetin	VAL 112	N	2.84	-7.03	7.51
	GLN 113	OE1	2.68		
	TRP 173	NE1	2.67		
	ARG 336	NH1	3.24		
	TYR 405	OH	2.70		
Eriodictyol	VAL 112	N	3.26	-6.84	7.38
	GLN 113	OE1	2.71		
	TRP 173	NE1	2.81		
	ARG 336	NH1	2.84		
Wogonin	TRP 173	NE1	2.88	-5.79	6.65
	ARG 336	NH1	2.92		
Chrysin	TRP 173	NE1	2.90	-5.37	5.86
	ARG 336	NH1	2.77		

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**Source of Support:** Nil. **Conflict of Interest:** None declared.