Anti-termite and antimicrobial efficacy of latexes from certain plant families

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Abstract

The present article explains the effect of plant latex and its formulations on termite control. Latex is natural plant polymer secreted by highly specialized cells known as laticifers. It is a milky white thick complex mixture of proteins, alkaloids, starch, sugars, oils, tannins, resins, and gums. Both latex and its components show multiple deleterious effects such as toxic, antifeedant, and repellent activities. These effectively inhibit growth and reproductive behavior in number of insect species. Latex components delay egg maturation, development, and inhibit gonad development in termites. Latex constituents display contact and systemic action and primarily used as poison baits to control soil termite. Latex-based combinatorial anti-termite formulations could be used in spray for termite control. The usage is safer than synthetic chemicals and will minimize the risk of poisoning of food chain, soil, and aqueous environment. This review article suggests the use of slow release of latex components inside soil when applied as poison baits for control of field termites.

Key words: Plant latex, Anti-termite activity, Latex secreting plant species, Latex component, Natural plant polymer

INTRODUCTION

ermites are one of the most agriculturally important insects and are known to cause enormous economic losses to many crop plants and tree species, buildings, etc. Termites are detritus feeders and feed on dead plants and trees. The main factors behind the presence of a large population of termites are humidity, mild temperature, and other climatic factors. The Indian white termite, Odontotermes obesus (Rambur) (Isoptera: Odontotermitidae), is highly destructive polyphagous insect pest, lives in huge mounds, and feeds on cellulose material and almost anything which contains carbohydrate. It causes economic damage to commercial wood, fibers, cellulose, sheets, papers, clothes, woolens and mats, and woody building material and infests green standing foliage, and cereals stored in go down. Both worker and soldier termites harm non seasoned commercial wood and its formed materials. Whether it is a rural area or an urban domestic site, termite menace is everywhere. Termites are known as silent destroyers because of their ability to chew wood, clothes, and harm to crop fields. Each year termites cause about \$ 30 billion in property damage.

There are about 2000 known termite species in the world. There are four types of termites

found in all over the world, that is, subterranean dry wood damps wood and powder post. Subterranean termites are highly dreadful damage about 95% of crop systems and other materials all over the world. In Tarai belt of Gorakhpur termites, menace is seen in different local regions, mainly in crop fields, household, and forests. In this area, the main species of termite are O. obesus (Indian white termite and red termite Coptotermes sp.). Subterranean termites are major pest worldwide, causing billion of loss in crops and household things annually. Similarly, at the global level in both tropical and sub-tropical countries, the infestation of drywood termite Cryptotermes brevis (Kalotermitidae) is one of the most important wood structural pests in the world.[1] Termites have presented human society with some of its greatest development challenges by consuming crops and damaging infrastructure. From report, these reported goods and services are a rough estimate that invasive insects cost a minimum of US\$30.0 billion per year globally[2] The subterranean termite Globitermus sulphureus are an important Southeast Asian pest causes damages to agriculture

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Received: 05-02-2022 **Revised:** 21-03-2022 **Accepted:** 29-03-2022 crops and building structures.^[3] Latexes are secreted by many flowering plant families. In this article, the latex and other plant natural products have been suggested for termite control. They inhibit metabolism in termites and kill them due to antifeedant, repellent, and toxic action.

Termites infest at various stages of plant growth and cause severe losses in sugarcane, maize, wheat, fruits, etc.^[4,5] In crop fields, termites cause 50-100% losses in yield. [6,7] Termite heavily damages plant foliage and destroys young saplings. However, for controlling termite attack, harmful synthetic chemical pesticides are extensively applied.[8] For controlling termite on crop plants, various synthetic pesticides such as cyclodiene, [9,10] hydroquinone, and indoxcarb[11] have been used. Dursban spray was found highly effective in the management of wood destroying termites.[12] These chemicals put serious deleterious effect on non-targeted biotic and abiotic factors of environment.[13] Although, chemical insecticides are highly effective against termite, they are hazardous to non-target organisms in the ecosystem.^[14] It's bound residues persists for longer duration in the environment, and through various trophic levels, they entered into the food chain. Hence, there is a need to replace chemical pesticides by natural plant origin pesticides.

SOURCE OF INFORMATION

For writing this comprehensive research review on plant latexes, various databases were searched. For the collection of relevant information, specific terms such as phytochemical subject headings (PhytoSH) and key text words, such as "Family Apocynaceae, Moracae, Companulaceae," "plant latexes," and "insecticidal activity," published till 2022 were used in MEDLINE. Most specially for retrieving all articles pertaining to the use of plant latexes, electronic bibliographic databases were searched and abstracts of published studies with relevant information on the plant latexes were collected. Furthermore, additional references were included through searching the references cited by the studies done on the present topic. Relevant terms were used individually and in combination to ensure an extensive literature search. For updating the information about a subject and incorporation of recent knowledge, relevant research articles, books, conferences proceedings, and public health organization survey reports were selected and collated based on the broader objective of the review. This was achieved by searching databases, including "SCOPUS, Web of Science, EMBASE, PubMed, PMC, Publon, Swiss-Prot, and Google searches." From this common methodology, discoveries and findings were identified and summarized in this final review.

LATEX SECRETING PLANT SPECIES

More than 12,000 plant species secrete latex. [15] The major producers of latex are *Hevea brasiliensis*, *Ficus benjamina*,

Campanula glomerata, [16] and Chelidonium (Papaveraceae).[17] Some common latex secreting plants from family Euphorbiaceae are H. brasiliensi, Euphorbia bicolor, Synadenium grantii, Sapium glandulosum, Jatropha gossypiifolia, Hura epitans, and oton urucurana. Major latex secreting plants from Moraceae are Ficus carica, Maclura tinctoria, Maclura pomifera, Brosimum gaudichaudii, Antiaris toxicaria, Artocarpus heterophyllus, and Dorstenia luamensis. Taraxacum koksaghyz, Scorzonera latifolia, Lactuca serriola, Parthenium argentatum, Solidago virgaurea, and Artemisia annua are common latex bearing plants from family Asteraceae. Cannabis sativa and Humulus lupulus are two common latex secreting plants from family Cannabaceae.[18] Plants secrete larger amounts of latex that is used to fight against herbivorous insect pests.^[19]

Latex is the milky sap that is produced by specialized cells known as laticifers of several plant species. This tissue is found distributed in the mass of parenchymatous cells throughout plant body. They contain numerous nuclei which lie embedded in the thin lining layer of protoplasm. This tissue consists of thick walled, greatly elongated and much branched ducts containing milky or yellowish-colored liquids. Latex cell is also called as "non-articulate latex ducts," these ducts are independent units which extend as branched structures for long distances in the plant body. Latex vessels are mostly found in many latex secreting plant families such as Papaveraceae, Compositae, Euphorbiaceae, and Moracesae.

Apocynaceae is one of the largest and important families in angiosperm. This family is famous for latex secreting plants, that is, Allamanda, Alstonia, Calotropis, Catharanthus, Cerbera, Dyera, Kopsia, Nerium, Plumeria, and Vallaris. [23,24] The common latex secreting plants of family Apocynaceae are Allamanda cathartica, Alstonia angustiloba, Calotropis procera, Catharanthus roseus, Cerbera floribunda, Dyera costulata, Nerium oleander, Plumeria alba, Vallaris glabra, Hancornia speciosa, Acokanthera oblongifolia, Apocynum cannabinum, Thevetia peruviana, Rauvolfia serpentine, Plumeria rubra, Tabernaemontana divaricata, and *Himatanthus drasticus*).^[25] The milkweed plant secretes white or milky thick latex rich in cardenolides, proteinase, alkaloids, terpenoids, steroids, flavonoids, glycosides, simple phenols, lactones, and hydrocarbons and other bioactive compounds.^[25] Cysteine proteases and chitin-related proteins and other proteins play major role in defending plants from herbivores.^[26] These ingredients are responsible for their antibacterial, antifungal, anthelmintic, cytotoxic, and insect repellent activities.^[27] They show synergism to prevent the plants from herbivory or infection.^[21] Campanulaceae are recognized by their white latex.

Family Euphorbiaceae is well known for latex secretion. *Euphorbia* species contain diverse phytochemicals such as terpenoids, flavonoids, and polyphenols, which constitute the secondary metabolites.^[28,29] *Euphorbia* latex is highly toxic, irritant, and strongly inhibit feeding in herbivorous insects.^[30]

The latex is the most valuable product obtained from *Euphorbia* species. *F. benjamina* and *H. brasiliensis* latex coagulation mechanisms deter termites from feeding.^[31] *Euphorbia* plants are believed to be a promising source of phytochemicals used in the pharmacy and food industries (Table 1).

TERMITE CONTROL BY PLANT LATEX

Plants synthesize so many secondary metabolites including latex which protect plants from physical damage caused by chewing herbivores insects pests.^[21,32,33] Latex exudes from plant parts after having an injury. Plant latex contains alkaloids mainly glycosides which heavily deter herbivorous insects and target insect pests effectively.^[34] Alkaloid glycosides also display strong anti-termitic activity.^[35] Latex also contains proteins which inhibit feeding behaviors in termites.^[17] It protects from wound injuries against insect bites and infection.

Plant latex is secreted to maintain plant defense against a diverse group of organisms. [36-38] Latex is secreted by 10% of plant families to prevent plants from chewing herbivory.^[21] Plant latex shows counterattack insect invasion. It can be used in environmental-friendly pest management of not only termites but also other insect pests. It may help to control pests and reduces harmful use of pesticides. Termites can be controlled by green pest management using clean cultivation, sanitation, biological and cultural control, least toxic chemical pesticides, and minimum use of chemicals and avoid killing of non-target species by spraying in target locations.[39] Plant latices with various formulations were found toxic when applied as contact or spray against termites. A high termite repellency and mortality are reported after direct or forced indirect exposure to the plant latexes. Plant latex-based formulations could be used for soil treatments to check menace of soil termites.^[40] Use of synthetic chemicals is harmful for non-target organisms, these persist for longer time in environment. Hence, there is a need to search for plantderived compounds as an alternative for termite control.^[41]

There are few prominent plant families which produce larger amount of latex are Euphorbiaceae, Asclepiadaceae, Moraceae, Caricaceae, Papaveraceae, Apocynaceae, Cannabaceae, and Asteraceae. [42,43] Few other families Loganiaceae and Rubiaceae are monoterpenoid indole alkaloids in latex (Table 2). [44]

Bio-organic Products from Latexes

Cardenolides

Plant latex is a potential source of bioactive compounds mainly mixture of proteins, carbohydrates, oils, and secondary metabolites. Toxic cardenolides (cardiac glycosides) are also secreted by milkweeds and deter herbivores from feeding. [45] Cardenolides are chief constituent of latex of family Apocynaceae members. [46] Both Apocynaceae and *Asclepediaceae* possess more than 300 cardenolides. [47,48]

Cardenolide mainly cardiac glycosides, that is, strophanthidin beta-D-glucomethylosido-D-alloside and beta-D-digitoxosido-D-alloside has been isolated from Moraceae plants. Cardenolides and its derivatives covenosigenin, glucopyranoside, and acospectoside play an important role as antifeedant in termites. [49,50] Various cardenolides have been isolated from *Pergularia tomentosa*, [51] *Calotropis gigantean*, [52] *Asclepias curassavica*, [53] and *Nerium indicum* showed high toxicity against termites. [54] Similarly, ischarin and ischaridin from *C. procera* found toxic against termoites, [55] while cerberin from *Cerbera odollam* is toxic to termites and causes significant mortality. [56]

Alkaloids

Indole alkaloids such as ervatamines and its derivatives are major components of the latex in latex secreting plants. [57,58] These exhibit good insecticidal activity against insects much similar to rotenone. [59,60] The alkaloids vinblastine and vincristine are bisindole alkaloids derived from coupling vindoline and catharanthine, monoterpenoid indole alkaloids produced exclusively by the Madagascar periwinkle (*C. roseus*). These are used for insect suppression of insect population. [61] Iridoids alkaloids exhibit anti-insect properties. These could be used to produce bioinsecticides. [62]

The alkaloid rauvomitorine A-I and C-9-methoxymethylene-sarpagine isolated from latex of *Rauvolfia vomitoria*, inhibit acetylcholinesterase inhibitory activities in termites.^[63] Alkaloids from *T. divaricata* named Taberniacins A and B show anti-termite efficacy.^[64] Few important alkaloids alkaloid tabernaines from *Tabernaemontana bufalina* latex; melotenine A and aspidosperma isolated from the latex of *Melodinus axillaris*, were found toxic to termites.^[28,65] Other alkaloids isolated from melotenuines A-E isolated from *Melodinus henryi have* anti-termite properties.^[66,67] Alkaloids isolated from *Melodinus tenuicaudatus*^[68] and *Leuconotis eugeniifolia* show antiplasmodial activities and toxic to termites (Table 2).^[69]

Peptidases

Plant latexes from family Apocynaceae contain peptidase which showed rich cysteine-protease activity. Cysteine proteases, such as ficin and bromelain, show toxicity against termites. [70] Cysteine peptidases are the most abundant enzymes in latex fluids. These also provide defense against phytopathogens. [71] Similarly, cysteine peptidases: Procerain and procerain B isolated from latex of *C. procera* also show strong of proteolytic activity. [72] The presence of enzymatic activities in latex is used to make potential resistance against phytopathogens and insects. [71] A rich amount of cysteine peptidases is also reported in latex from *T. peruviana*. It also contains peptidase inhibitor, cysteine peptidases, peroxidases, and osmotin. [73]

Philibertain, caricain, [74] and asclepain [75] are cysteine peptidases inhibitor. Latex-bearing plants also host insects. Latex peptidase inhibitors also compete with inhibitors of

Tripathi and Upadhyay: Anti-termite activity of plant latex

	Table 1: Anti-termite activity of different plants latexes.				
S. N.	Name of plants	Common Name	Activity against species		
1.	Ficus lacor	Java fig	Anti-antiarthritic, antidiabetic, anti-inflammatory		
2.	Ficus hirta	Hairy mountain fig	Antibacterial and anti-cancerous		
3.	Ficus sarmentosa	Nepal fig	Antibacterial, antifungal, antioxidant activity		
4.	Ficus palmate	Jungli anjir	Antimicrobial, antibacterial activity.		
5.	Ficus neriifolia	Willow -leaf fig	Anti-inflammatory effects of phenolic compounds		
6.	Ficus semicordata	Drooping fig	Antioxidative and antibacterial activities		
7.	Ficus pumila	Creeping fig	Cause phytophotodermatitis potentially serious skin inflammation		
8.	Ficus virens	White fig	Anti-inflammatory activity		
9.	Milicia excels	African Teak	In vitro antioxidant staphylococcal activity		
10.	Ficus racemosa	Goolar	Various diseases/disorders including diabetes, liver disorders, diarrhea, inflammatory conditions, hemorrhoids, respiratory, and urinary diseases		
11.	Ficus religosa	Pippal	Treatment of pain, inflammation, impotence, menstrual disturbances, and urine related problems, and as uterine tonic		
12.	Ficus benghalensis	Banyan	Used for the treatment of neuralgia, rheumatism, lumbago, bruise, nasitis, gonorrhea, inflammations, cracks of the sole and skin disease and in Ayurveda for diarrhea, dysentery, and piles		
13.	Morus alba	White mulberry	Insecticidal activity		
14.	Ficus auriculata	Elephant ear fig	Anti-bacterial and antioxidant activity		
15.	Ficus carica	Common fig	Inhibition of cancer cell growth in digestive tract		
16.	Ficus hispida	Devil fig	Anti-diarrheal activity		
17.	Ficus elastic	Rubber fig	Anthelmintic activity and parasitic worm infection etc.		
18.	Ficus ampilissima	Indian Bat fig	Antibacterial, antifungal, antioxidant activity		
19.	Artocarpus heterophyllus	Jackfruit	Anti-inflammatory effects of phenolic compounds		
20.	Ficus sycomorus	Sycamore fig	Antibacterial, anti-inflammatory		

Table 2: Anti-termite activities of plant latexes from different family				
S. No.	Name of plant	Common Name	Activity against species	
1	Hevea brasiliensis	Rubber tree	Coagulation mechanisms among the more than 20,000 Latex-bearing plant species are lacking	
2	Chelidonium majus	Greater celandine	Traditional folk medicine to treat papillae, warts, condylomas, which are visible effects of human papilloma virus infections	
3	E. peplus	Radium weed	Constitutive defense metabolites against insect herbivores and pathogens for the plant	
4	Thevetia peruviana	Yellow oleander	Antifungal activity against the isolates followed by Manilkara zapota	
5	Р. атара	Amapa-Amargoso	Change Chrysomya megacephala post-embryonic development	
6	Plumeria pudica	Golden Arrow	Animals against inflammatory ulcerative colitis	
7	Synadenium grantii	African milk bush	Nematicidal activity on Meloidogyne incognita and Panagrellus redivivus	
8	Euphorbia obtusifolia	Spurge	Inhibitory activity on the mammalian mitochondrial	
9	Euphorbia tirucalli	Indian tree spurge	Determine the molecular basis of the laticifers functions in this plant	
10	Plumeria rubra	White Frangipani	Against Aedes aegypti and Anopheles stephensi	

gut peptidases and show resistance susceptibility of plant latex to termites. [76] Araujiain is latex cystein peptidases from *Araujia angustifolia* shows proteolytic activity and can be used as a potent termicidal agent to kill the symbionts of termites. [77] Cystein peptidases ervatamin-A, ervatamin-B, and ervatamin-C from *Ervatamia coronaria* show anti-termitic

activity (Ghosh *et al.*, 2008).^[78] Cysteine peptidases named 12, 16-dihydroxicalotropin, calotropin, corotoxigenin 3-O-glucopyranoside, and desglucouzarin isolated from the latex of *Asclepias subulata*, it promotes cell death through caspase-dependent apoptosis in termites. Plant latex acts as an anticoagulant to stop bleeding and wound healing.^[79]

Cysteine proteases from family Apocynaceae plants latex exhibited both thrombin and plasmin like activities.

Flavonoids

Flavonoids are secondary metabolites having a polyphenolic structure. These are widely found in fruits, vegetables, and certain beverages.[80] Biochanin A flavonoids from Apocynaceae plant latex found most effective in reducing fecundity and it also acts as antifeedant against Coptotermes formosanus Shiraki.[81] The chief flavonoids from the latex of Apocynum venetum are plumbocatechin A, 8-O-methylretusin and kaempferol 3-O-(6"-O-acetyl)-β-D-galactopyranoside. These easily kill termite gut symbionts that result in death of termites.[82] Similarly, flavonoids kaemperol-3-Orutinoside, quercetin-3-O-glucoside, and kaemperol-3-O-glucoside isolated from Holarrhena floribunda show antioxidant activity.[83] Flavonoids named kaempferol 3-rhamnogluco-7-glucoside, kaempferol 3-rhamnogluco-7galactoside, quercetin 3-rutino-7-glucoside, and quercetin 3-rhamnogluco-7-glucoside from the latex Vinca minor termicidal activity.[84] Flavonoids naringenin, aromadendrin (dihydrokaempferol), and kaempferol are chief constituents of Echites hirsute latex, they show termicidal properties[85] and flavonoids isolated from Trachelospermum jasminoides named apigenin, apigenin 7-O-beta-glucoside, apigenin 7-O-beta-neospheroside, naringin, and 6,8-di-Cglucopyanosylapigenin show antifeedant activity.[86]

Terpenes

Like alkaloids, the abundance of terpenes and their derivatives has been reported in many members of the family Apocynaceae. Terpenoids isolated from N. oleander are mainly oleandric acid, ursolic acid, betulinic acid, betulin, and derivatives of epoxydammarane 3\beta, 25-diol show significant anti-termitic activity.[87] Among the various isolated terpenes, vulgarone B, apiol, and enicin exhibit significantly higher mortalities in termites. These are highly toxic compounds to termites.^[88] The terpenoids constitute the largest class of natural products and many interesting products are extensively applied in insect pest management such as termicides.^[89] Terpenes ursolic acid isolated from Pleiocarpa pycnantha show toxic against termites. [90] Pentalinon andrieuxii latex contains urechitol A terpenes show antifeedant activity.[91] Major terpenes perisomalien A, lupeol acetate, β -amyrin cycloart-23Z-ene-3 β , 25-diol, and β -sitosterol-3-O- β -Dglucopyranoside isolated from Periploca somaliensis latex showed toxic effects against termites. [92]

Sterols

Stigmasterol, β -sitosterol, and campesterol are phytosterols or steroid alcohols. These show play important role in insect growth and development and its scarcity seize them at any stage of life including larval to adults forms. Sterols show strong insecticidal activities against adult termites and a negative impact on fecundity. [93] Latex of *Gymnema sylvestre*

contains beta-sitosterol, campesterol, and stigmasterol. These show antifeedant activity against termites. [94] Sterols pentalinonside and pentalinonsterol isolated from *P. andrieuxii* show termicidal activity. [95] *Alstonia scholaris* latex contains sterols named poriferasterol, epicampesterol, β -sitosterol, β -sitosterol, showanti-termitic activities. [96] β -sitosterol and β -daucosterol are chief sterols found in the latex of *Periploca forrestii*, they are highly toxic to termites. In the latex of *P. tomentosa*, 3-acetyltaraxasterol, 3-taraxasterol, and 16α -hydroxytaraxasterol-3-acetate are chief sterols which are responsible for the significant mortality in termites. [97] Similarly, *Cynanchum limprichtii* latex contains limproside A and limproside B which are proven to be toxic against termites. [98]

Simple phenolic compounds

The phenolic compounds show insecticidal activity and use in insect control because of their strong action on insect digestion. Phenolic compound ellagic acid derivatives (Shi *et al.*, 2010), protocatechuic acid, catechin, and quercetin isolated from *H. speciosa* showed deterrent and insecticidal activity against termites. P-coumaric and ferulic acid both are phenolic compounds found in *Asclepias linaria* latex showed toxic effect against termites. The phenolic compound present in the latex of *Cynanchum wilfordii* is $2-O-\beta$ -laminaribiosyl-4-hydroxyacetophenone responsible for significant mortality in termites.

Lignans

Lignan glycoside, (+)-pinoresinol 4-O-[6"-O-vanilloyl]-β-d-glucopyranoside isolated from the latex of *C. gigantea* show insecticidal activity. Similar activity is also reported in reveled carbinol as phenolic lignan found in *Carissa carandas* and *Carissa carandas*. Recently, syringaresinol 4-O-b-glucopyranoside isolated from *Vinca major* is an insect deterrent. [105]

Insecticidal Activity of Latex

Various plants belongs to moraceae family such as Ficus lacor, Ficus hirta, Ficus sarmentosa, Ficus palmate, Ficus neriifolia, Ficus semicordata, Ficus pumila, Ficus virens, Milicia excels, Ficus racemosa, Ficus religosa, Ficus benghalensis, Morus alba, Ficus auriculata, Ficus carica, Ficus elastic, Ficus ampilissima, Artocarpus heterophyllus and Ficus sycomorus have different potential like Antibacterial, antifungal, antioxidant, Termiticidal, Antimicrobial, antibacterial activity, Anti antiarthritic, antidiabetic and anti inflammatory activities (Table 1). Latexes from different family plants like Hevea brasiliensis ,Chelidonium majus,E. peplus,Thevetia peruviana, P. amapa ,Plumeria pudica, Synadenium grantii, Euphorbia obtusifolia, Euphorbia tirucalli, Plumeria rubra showed high potential as antifungal, antioxidant, Termiticidal, Antimicrobial and antibacterial activity (Table 2).

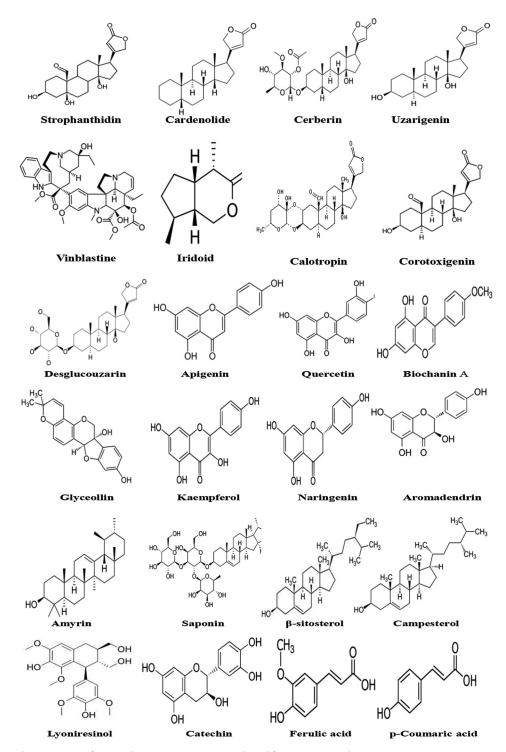


Figure 1: Chemical structures of major latex components isolated from various plant species

Latex components if used as insecticide are safe and do not put any adverse effects on environmental quality and non-target organisms including human health. [106] Latex ingredients are bioactive substances and are useful as medicines and pesticides. [107] Plant latex provides an alternative way in the insect control strategy. These are secondary chemicals synthesized by plants [108] which are non-toxic and biodegradable and can be used to control insects as

an alternative to chemical insecticide. [109] The latex from the different plant families used since long time in medicine as well as in the control of some insects. [110] Latex fractioned with different solvents shows a variable degree of antitermitic activities. In the latex of *Euphorbia kansui*, there are six triterpenoids, one of them is identifies as euphane-3 β and 20-dihydroxy-24-ene (Figure 1). All these compounds have insecticidal activity. [111]

A cysteine protease isolated from seeds of *Albizia procera* ApCP encapsulated with graphene quantum dots showed enhanced toxicity against insects. [112] Most of protease activity is seen toxic to insect midgut and to the cuticle). [113] The leaf *Aloe* sp. latex is found active against microbes and insects. [114] Latex contains highly active anti-termite compounds which showed high toxicity against insects. [115] Plant latex contains secondary metabolites help the plant in making defense against insect pests. Latex components act antifeedant [116] and act as insect growth control agents. The insecticidal activities of latex are dose dependent. [117]

Latex from Morus alba contains protein a and b (LA-a and LA-b) which show significant chitinase and chitosanase activities. LA proteins hydrolyze chitin surface of insects. [118] Lobelia siphilitica shows reduced latex production and high herbivores attack.[119] Carica papaya latex solvent extracts showed larvicidal properties against a number of insects.[117] Plants also bear morphological structures, that is, waxes, trichomes, and latices make the feeding more difficult for the insects.^[120] The latex from the leaves of *Aloe trigonantha* is a sticky poisonous exudates that make insect mouth parts functionless.[21] It acts as feeding deterrent. The latex of Garcinia morella (Gaertn.) possesses 5-Oxohexanenitrile (18.7%), phenol, 2, 4-bis (1, 1-dimethylethyl) (24.64%), and hexadecanoic acid (22.85%) (Figure 1). These latex compounds show toxicity against many insects.[121] In cucurbit plants, phloem latex exudates from phloem from cut sieve tubes used to make defense against herbivores.[122] Latex exudates stop development of first and third instar larvae enclosed on Lobelia cardinalis (Campanulaceae) failed to develop.

ANTIMICROBIAL ACTIVITY

Besides insecticidal activity plant latex component also showed strong antimicrobial effects. A latex component aloesin is a C-glycosylated chromone, it exhibits antibacterial activity against pathogens.[123] The leaf latex of Amanita citrina shows antimalarial activity. Hevea latex allergy shows IgE-mediated allergy. C. majus (Papaveraceae) latex found active against bacteria, fungi, viruses, protozoans, nematodes, and insects. It also shows some anti-cancer properties. [124] Iridoid alkaloids possess good to excellent activities against phytopathogenic fungi Fusarium graminearum. Euphorbia plant latex shows antimicrobial activity but its exposure is harmful for humans' microbes. Euphorbiaceae plant latexes showed string antimicrobial activity. It is also used for making traditional medicine to combat microbial infections. Chrysanthemum and Uniflower Swisscentaury root extracts showed anti-angiogenic effects in zebrafish. Latex contains hydrolytic active proteins which also work much similar to proteases.[42] The constituents of latex are well known in some plant families for their phytotoxic, insecticidal, cytotoxic, antibacterial, and antifungal activities.

CONCLUSION

It is clear from various studies that plant latex contains deterrent chemicals which elicit the feeding behavior in insects. These latex-based pure compounds are quite effective and might have potential uses in agriculture mainly for termite control. These inhibit feeding in first to third instar larvae; inhibit its growth and molting. These effectively inhibit embryonic and post-embryonic development in insects. Plant latex contains highly active bio-organic components mainly glycosides, hydrolytic active proteins protease inhibitors, sterols, phenylpropanoids, monoterpenes, and furanocoumarins and Lactucin (*Lactuca sativa*), myristicin, are antagonist of nicotinic acetylcholine receptors found in insects. Most of them are hemotoxic, cytotoxic, and neurotoxic to insects. Among them, proteases can be used as potential pesticides in place of synthetic insecticides. Lignan glycoside, isolated from the latex of C. gigantea, shows insecticidal activity. Both latex and its derived formulations cause significant mortality in termites. These natural formulations put no deleterious effect on non-targeted biotic and abiotic factors of environment. These easily biodegrade in the medium and show no residual effect and non-harmful to food chain. These formulations can be used as dust, spray, and in form poison baits to control termite population in field garden and vegetation, particularly in semi-arid ecosystems.

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CONFLICTS OF INTEREST

The author declares no conflicts of interest regarding the publication of this paper.

REFERENCES

- Santos AA, de Oliveira BM, Melo CR, Lima AP, Santana ED, Blank AF, et al. Sub-lethal effects of essential oil of Lippia sidoides on drywood termite Cryptotermes brevis (Blattodea: Termitoidea) Ecotoxicol Environ Saf 2017;145:436-41.
- 2. Bradshaw CJ, Leroy B, Bellard C, Roiz D, Albert C, Fournier A, *et al.* Massive yet grossly underestimated global costs of invasive insects. Nat Commun 2016;7:12986.
- 3. Hussin NA, Najimudin N, Ab Majid AH. The *de novo* transcriptome of workers head of the higher group termite *Globitermes sulphureus* Haviland (*Blattodea: Termitidae*). Heliyon 2019;5:e02969.
- 4. Salihah Z, Satar A, Khatoon R. Termite Damage to Sugarcane Crop at Mardan Area. Ann technology Report. Tarnab, Peshawar, Pakistan: NIFA; 1986. p. 126-9.

- Sattar A, Salihah Z. Detection and Control of Subterranean Termites. Technologies for Sustainable Agriculture. Proceedings National Workshop (Technologies for Sustainable Agriculture). Faisalabad, Pakistan: NIAB; 2001. p. 195-8.
- Rao MR, Singh MP, Day R. Insect pest problem in tropical agroforestry systems, contributory factors and strategie for management. Agroforestry Syst 2000;50:243-77.
- Sekamatte MB, Kyamanywa S, Wilson HR, Simth AR.
 The effect of placement method and rate of application of crushed bones on the activity of predatory ants and their impact on termite damage to maize. Insect Sci Appl 2002;22:199-204.
- Venkateswara Rao J, Parvathi K, Kavitha P, Jakka NM, Pallela R. Effect of chlorpyrifos and monocrotophos on locomotor behaviour and acetylcholinesterase activity of subterranean termites, *Odontotermes obesus*. Pest Manag Sci 2005;61:417-21.
- Sim M, Forbes A, McNeil J, Robert G, Termite control and other determinants of high body burdens of cyclodiene insecticides. Arch Environ Health 1998;53:114-23.
- Sisay A, Ibrahim A, Tefera T. Management of Termite (Microtermes adschaggae) on hot pepper using powdered leaves and seeds of some plant species at Bako, Western Ethiopia. East Afr J Sci 2008;2:40363.
- Hu XP, Song D, Scherer CW. Transfer of indoxacarb among workers of *Coptotermes formosanus (Isoptera: Rhinotermitidae*): Effects of dose, donor: Recipient ratio and post-exposure time. Pest Manag Sci 2005;61:1209-14.
- 12. Roll D. In: Randall CJ, editor. Management of Wood-destroying Pests. Ohio:2007. p. 8-73. 2007.
- 13. Pimental D. Amounts of pesticides reaching target pests: Environmental impacts and ethics. J Agric Environ Ethics 1995;8:17-29.
- 14. Kumar R, Nitharwal M, Chauhan R, Pal V, Kranthi KR. Evaluation of eco-friendly control methods for management of mealybug, *Phenacoccus solenopsis* Tinsley in cotton. J Entomol 2012;9:32-40.
- 15. Hagel JM, Yeung EC, Facchini PJ. Got milk? The secret life of laticifers. Trends Plant Sci 2008;13:631-9.
- Bauer G, Gorb SN, Klein MC, Nellesen A, von Tapavicza M, Speck T. Comparative study on plant latex particles and latex coagulation in *Ficus benjamina*, *Campanula glomerata* and three *Euphorbia* species. PLoS One 2014;9:e113336.
- 17. Nawrot R. Defense-related proteins from *Chelidonium majus* L. as important components of its latex. Curr Protein Pept Sci 2017;18:864-80.
- 18. Metcalfe CR. Distribution of latex in the plant kingdom. Econ Bot 1967;21:115-27.
- 19. Tasca JA, Smith CR, Burzynski EA, Sundberg BN, Lagalante AF, Livshultz T, *et al.* HPLC-MS detection of pyrrolizidine alkaloids and their *N*-oxides in herbarium specimens dating back to the 1850s. Appl Plant Sci 2018;6:e1143.
- 20. Ramos MV, Freitas CD, Stanisçuaski F, Macedo LL,

- Sales MP, Souza DP, *et al.* Performance of distinct crop pests reared on diets enriched with latex proteins from *Calotropis procera*: Role of laticifer proteins in plant defense. Plant Sci 2007;173:349-57.
- 21. Dussourd DE, Van Valkenburg M, Rajan K, Wagner DL. A notodontid novelty: *Theroa zethus* caterpillars use behavior and anti-predator weaponry to disarm host plants. PLoS One 2019;14:e0218994.
- 22. Rosa SS, Rosa MF, Fonseca MA, Luz GV, Avila CF, Domínguez AG, *et al.* Evidence in practice of tissue healing with latex biomembrane: Integrative review. J Diabetes Res 2019;2019:7457295.
- 23. Chan EW, Wong SK, Chan HT. Apocynaceae species with antiproliferative and/or antiplasmodial properties: A review of ten genera. J Integr Med 2016;14:269-84.
- 24. Cristina RT, Dumitrescu E, Brezovan D, Muselin F, Chiurciu V. Effect of *Euphorbia cyparissias* ointments on acanthosis. Afr J Tradit Complement Altern Med 2014;11:1-6.
- 25. Bhadane BS, Patil MP, Maheshwari VL, Patil RH. Ethnopharmacology, phytochemistry, and biotechnological advances of family *Apocynaceae*: A review. Phytother Res 2018;32:1181-210.
- 26. Konno K. Plant latex and other exudates as plant defense systems: Roles of various defense chemicals and proteins contained therein. Phytochemistry 2011;72:1510-30.
- 27. Salomé Abarca LF, Klinkhamer PG, Choi YH. Plant latex, from ecological interests to bioactive chemical resources. Planta Med 2019;85:856-68.
- 28. Shi BB, Chen J, Bao MF, Zeng Y, Cai XH. Alkaloids isolated from *Tabernaemontana bufalina* display xanthine oxidase inhibitory activity. Phytochemistry 2019;166:112060.
- 29. Hua J, Liu Y, Xiao CJ, Jing SX, Luo SH, Li SH. Chemical profile and defensive function of the latex of *Euphorbia peplus*. Phytochemistry 2017;136:56-64.
- 30. Machado MM, de Oliveira LF, Zuravski L, de Souza RO, Fischer P, Duarte JA, *et al.* Evaluation of genotoxic and cytotoxic e_ects of hydroalcoholic extract of *Euphorbia tirucalli* (*Euphorbiaceae*) in cell cultures of human leukocytes. An Acad Bras Cienc 2016;88:17-28.
- 31. Bauer G, Friedrich C, Gillig C, Vollrath F, Speck T, Holland C. Investigating the rheological properties of native plant latex. J R Soc Interface 2013;11:0847.
- 32. Huber M, Epping J, Schulze Gronover C, Fricke J, Aziz Z, Brillatz T, *et al.* A latex metabolite benefits plant fitness under root herbivore attack. PLoS Biol 2016;14:e1002332.
- 33. Ernst M, Nothias LF, van der Hooft JJ, Silva RR, Saslis-Lagoudakis CH, Grace OM, *et al.* Assessing specialized metabolite diversity in the cosmopolitan plant genus *Euphorbia* L. Front Plant Sci 2019;10:846.
- 34. Runguphan W, Maresh JJ, O'Connor SE. Silencing of tryptamine biosynthesis for production of nonnatural alkaloids in plant culture. Proc Natl Acad Sci U S A 2009;106:13673-8.

- 35. Araya JJ, Binns F, Kindscher K, Timmermann BN. Verticillosides A-M: Polyoxygenated pregnane glycosides from *Asclepias verticillata* L. Phytochemistry 2012;78:179-89.
- 36. Agrawal AA, Hastings AP. Plant defense by latex: Ecological genetics of inducibility in the milkweeds and a general review of mechanisms, evolution, and implications for agriculture. J Chem Ecol 2019;45:1004-18.
- 37. Sibi G, Wadhavan R, Singh S, Shukla A, Dhananjaya K, Ravikumar KR, *et al.* Plant latex: a promising antifungal agent for post harvest disease control. Pak J Biol Sci 2013;16:1737-43.
- 38. Livshultz T, Kaltenegger E, Straub SC, Weitemier K, Hirsch E, Koval K, *et al.* Evolution of pyrrolizidine alkaloid biosynthesis in Apocynaceae: Revisiting the defence de-escalation hypothesis. New Phytol 2018;218:762-73.
- 39. Maddala VK. Green pest management practices for sustainable buildings: Itical review. Sci Prog 2019;102:141-52.
- 40. Bläske VU, Hertel H. Repellent and toxic effects of plant extracts on subterranean termites (*Isoptera*: *Rhinotermitidae*). J Econ Entomol 2001;94:1200-8.
- 41. Xie Y, Huang Q, Lei C. Bioassay-guided isolation and identification of antitermitic active compound from the leaf of Chinese cedar (*Cryptomeria fortunei* Hooibrenk). Nat Prod Res 2013;27:2137-9.
- 42. Sytwala S, Domsalla A, Melzig MF. Investigation of plant latices of *Asteraceae* and Campanulaceae regarding proteolytic activity. Plant Physiol Biochem 2015;97:117-23.
- 43. Lewinsohn T. The geographical distribution of plant latex. Chemoecology 1991;2:64-8.
- 44. Szabó LF. Diversity and selectivity in biomolecules. Acta Pharm Hung 2001;71:392-404.
- 45. Rasmann S, Agrawal AA. Latitudinal patterns in plant defense: Evolution of cardenolides, their toxicity and induction following herbivory. Ecol Lett 2011;14:476-83.
- Agrawal AA, Petschenka G, Bingham RA, Weber MG, Rasmann S. Toxic cardenolides: Chemical ecology and coevolution of specialized plant-herbivore interactions. New Phytol 2012;194:28-45.
- 47. Wen S, Chen Y, Lu Y, Wang Y, Ding L, Jiang M. Cardenolides from the *Apocynaceae* family and their anticancer activity. Fitoterapia 2016;112:74-84.
- 48. Züst T, Petschenka G, Hastings AP, Agrawal AA. Toxicity of milkweed leaves and latex: Chromatographic quantification versus biological activity of cardenolides in 16 *Asclepias* species. J Chem Ecol 2019;45:50-60.
- Dobler S, Petschenka G, Pankoke H. Coping with toxic plant compounds--the insect's perspective on iridoid glycosides and cardenolides. Phytochemistry 2011;72:1593-604.
- 50. Pecio Ł, Hassan EM, Omer EA, Gajek G, Kontek R, Sobieraj A, *et al.* Cytotoxic Cardenolides from the Leaves of *Acokanthera oblongifolia*. Planta Med

- 2019;85:965-72.
- 51. Piacente S, Masullo M, De Nève N, Dewelle J, Hamed A, Kiss R, *et al.* Cardenolides from *Pergularia tomentosa* display cytotoxic activity resulting from their potent inhibition of Na+/K+-ATPase. J Nat Prod 2009;72:1087-91.
- 52. Seeka C, Sutthivaiyakit S. Cytotoxic cardenolides from the leaves of *Calotropis gigantea*. Chem Pharm Bull (Tokyo) 2010;58:725-8.
- 53. Li JZ, Qing C, Chen CX, Hao XJ, Liu HY. Cytotoxicity of cardenolides and cardenolide glycosides from *Asclepias curassavica*. Bioorg Med Chem Lett 2009;19:1956-9.
- 54. Wang XB, Li GH, Zheng LJ, Ji KY, Lü H, Liu FF, *et al.* Nematicidal cardenolides from *Nerium indicum* Mill. Chem Biodiv 2009:6:431-6.
- 55. Sweidan NI, Abu Zarga MH. Two novel cardenolides from *Calotropis procera*. J Asian Nat Prod Res 2015;17:900-7.
- 56. Menezes RG, Usman MS, Hussain SA, Madadin M, Siddiqi TJ, Fatima H, *et al. Cerbera odollam* toxicity: A review. J Forensic Leg Med 2018;58:113-6.
- 57. Castillo L, Rossini C. Bignoniaceae metabolites as semiochemicals. Molecules 2010;15:7090-105.
- 58. Zhang Y, Wang H, Tao L, Huang AX. Milk-clotting mechanism of *Dregea sinensis* Hemsl. Protease. J Dairy Sci 2015;98:8445-53.
- 59. Rosales PF, Bordin GS, Gower AE, Moura S. Indole alkaloids: 2012 until now, highlighting the new chemical structures and biological activities. Fitoterapia 2020;143:104558.
- Xia Q, Tian H, Li Y, Yu X, Zhang W, Wang Q. Biomimetic synthesis of iridoid alkaloids as novel leads for fungicidal and insecticidal agents. J Agric Food Chem 2020;68:12577-84.
- 61. Liscombe DK, O'Connor SE. A virus-induced gene silencing approach to understanding alkaloid metabolism in *Catharanthus roseus*. Phytochemistry 2011;72:1969-77.
- Chowański S, Adamski Z, Marciniak P, Rosiński G, Büyükgüzel E, Büyükgüzel K, et al. A review of bioinsecticidal activity of solanaceae alkaloids. Toxins (Basel) 2016;8:60.
- 63. Zhan G, Miao R, Zhang F, Wang X, Zhang X, Guo Z. Cytotoxic yohimbine-type alkaloids from the leaves of *Rauvolfia vomitoria*. Chem Biodiv 2020;17:e2000647.
- 64. Hirasawa Y, Tougan T, Horii T, Hadi AHA, Morita H. Leucophyllinines A and B, bisindole alkaloids from *Leuconotis eugeniifolia*. J Nat Med 2019;73:533-40.
- 65. Fang L, He TT, Zhu KK, Zhang M, Zhou J, Zhang H. Cytotoxic aspidosperma-type alkaloids from *Melodinus axillaris*. J Asian Nat Prod Res 2019;21:284-90.
- 66. Ma K, Wang JS, Luo J, Kong LY. Six new alkaloids from *Melodinus henryi*. Fitoterapiam 2015;100:133-8.
- 67. He J, Zhang FL, Li ZH, Yang HX, Shao Q, Feng T, et al. Monoterpenoid indole alkaloids from the bark of *Melodinus henryi*. Fitoterapia 2019;138:104354.
- 68. Liu JW, Huo ZQ, Zhao Q, Hao XJ, He HP, Zhang Y.

- Melotenuines A-E, cytotoxic monoterpenoid indole alkaloids from *Melodinus tenuicaudatus*. Fitoterapia 2019;138:104347.
- 69. Tang Y, Nugroho AE, Hirasawa Y, Tougan T, Horii T, Hadi AH, *et al.* Leucophyllinines A and B, bisindole alkaloids from *Leuconotis eugeniifolia*. J Nat Med 2019;73:533-40.
- 70. Konno K, Hirayama C, Nakamura M, Tateishi K, Tamura Y, Hattori M, *et al.* Papain protects papaya trees from herbivorous insects: Role of cysteine proteases in latex. Plant J 2004;37:370-8.
- 71. Freitas CD, Silva RO, Ramos MV, Porfirio CT, Farias DF, Sousa JS, *et al.* Identification, characterization, and antifungal activity of cysteine peptidases from *Calotropis procera* latex. Phytochemistry 2020;169:112163.
- Ramos MV, Araújo ES, Jucá TL, Monteiro-Moreira AC, Vasconcelos IM, Moreira RA, et al. New insights into the complex mixture of latex cysteine peptidases in Calotropis procera. Int J Biol Macromol 2013;58:211-9.
- 73. De Freitas CD, da Cruz WT, Silva MZ, Vasconcelos IM, Moreno FB, Moreira RA, *et al.* Proteomic analysis and purification of an unusual germin-like protein with proteolytic activity in the latex of *Thevetia peruviana*. Planta 2016;243:1115-28.
- 74. Torres MJ, Natalucci C, López LM, Trejo SA. Insights into the hydrolytic activity of *Asclepias fruticosa* L. protease. Biotechnol Lett 2019;41:1043-50.
- 75. Sequeiros C, Torres MJ, Trejo SA, Esteves JL, Natalucci CL, López LM. Philibertain g I, the most basic cysteine endopeptidase purified from the latex of *Philibertia gilliesii* (*Apocynaceae*). Protein J 2005;24:445-53.
- 76. Ramos MV, Pereira DA, Souza DP, Silva ML, Alencar LM, Sousa JS, *et al.* Peptidases and peptidase inhibitors in gut of caterpillars and in the latex of their host plants. Planta 2015;241:167-78.
- 77. Obregón WD, Lufrano D, Liggieri CS, Trejo SA, Vairo-Cavalli SE, Avilés FX, *et al.* Biochemical characterization, cDNA cloning, and molecular modeling of araujiain aII, a papain-like cysteine protease from *Araujia angustifolia* latex. Planta 2011;234:293-304.
- 78. Ghosh R, Chakraborty S, Chakrabarti C, Dattagupta JK, Biswas S. Structural insights into the substrate specificity and activity of ervatamins, the papain-like cysteine proteases from a tropical plant, *Ervatamia coronaria*. FEBS J 2008;275:421-34.
- Shivaprasad HV, Rajesh R, Nanda BL, Dharmappa KK, Vishwanath BS. Thrombin like activity of *Asclepias curassavica* L. latex: Action of cysteine proteases. J Ethnopharmacol 2009;123:106-9.
- 80. Panche A, Diwan A, Chandra S. Flavonoids: An overview. J Nutr Sci 2016;5:E47.
- 81. Boué SM, Raina AK. Effects of plant flavonoids on fecundity, survival, and feeding of the Formosan subterranean termite. J Chem Ecol 2003;29:2575-84.
- 82. Kong NN, Fang ST, Liu Y, Wang JH, Yang CY, Xia CH. Flavonoids from the halophyte *Apocynum venetum* and their antifouling activities against marine

- biofilm-derived bacteria. Nat Prod Res 2014;28:928-31.
- 83. Badmus JA, Ekpo OE, Rautenbach F, Marnewick JL, Hussein AA, Hiss DC. Isolation and antioxidant activity of flavonoids from *Holarrhena floribunda* (G.don) leaves. Acta Biochim Pol 2016;63:353-8.
- 84. Szostak H, Kowalewski Z. The flavonoids in the leaves of *Vinca minor* L. (*Apocynaceae*). Pol J Pharmacol Pharm 1975;27:657-63.
- 85. Chien MM, Svoboda GH, Schiff PL Jr., Slatkin DJ, Knapp JE. Chemical constituents of *Echites hirsuta* (*Apocynaceae*). J Pharm Sci 1979;68:247-9.
- 86. Tan XQ, Guo LJ, Chen HS, Wu LS, Kong FF. Study on the flavonoids constituents of *Trachelospermum jasminoides*. Zhong Yao Cai 2010;33:58-60.
- 87. Fu L, Zhang S, Li N, Wang J, Zhao M, Sakai J, *et al.* Three new triterpenes from *Nerium oleander* and biological activity of the isolated compounds. J Nat Prod 2005;68:198-206.
- 88. Meepagala KM, Osbrink W, Burandt C, Lax A, Duke SO. Natural-product-based chromenes as a novel class of potential termiticides. Pest Manag Sci 2011:67:1446-50.
- 89. Singh B, Sharma RA. Plant terpenes: Defense responses, phylogenetic analysis, regulation and clinical applications. 3 Biotech 2015;5:129-51.
- 90. Omoyeni OA, Meyer M, Iwuoha E, Green I, Hussein AA. An unusual 2, 3-secotaraxerene and other cytotoxic triterpenoids from *Pleiocarpa pycnantha* (*Apocynaceae*) leaves collected from Nigeria. Molecules 2014;19:3389-400.
- 91. Hiebert-Giesbrecht MR, Escalante-Erosa F, García-Sosa K, Dzib GR, Calvo-Irabien LM, Peña-Rodríguez LM. Spatio-temporal variation of terpenoids in wild plants of *Pentalinon andrieuxii*. Chem Biodiv 2016;13:1521-6.
- 92. Jabal KA, Abdallah HM, Mohamed GA, Shehata IA, Alfaifi MY, Elbehairi SE, *et al.* Perisomalien A, a new cytotoxic scalarane sesterterpene from the fruits of *Periploca somaliensis*. Nat Prod Res 2020;34:2167-72.
- 93. Beaulieu R, Grand E, Stasik I, Attoumbré J, Chesnais Q, Gobert V, *et al.* Synthesis and insecticidal activities of novel solanidine derivatives. Pest Manag Sci 2019;75:793-800.
- 94. Vats S, Kamal R. *In vivo* and *in vitro* evaluation of sterols from *Gymnema sylvestrte* R. Br. Pak J Biol Sci 2013;16:1771-5.
- 95. Pan L, Lezama-Davila CM, Isaac-Marquez AP, Calomeni EP, Fuchs JR, Satoskar AR, *et al.* Sterols with antileishmanial activity isolated from the roots of *Pentalinon andrieuxii*. Phytochemistry 2012;82:128-35.
- Wang CM, Yeh KL, Tsai SJ, Jhan YL, Chou CH. Antiproliferative activity of triterpenoids and sterols isolated from *Alstonia scholaris* against non-small-cell lung carcinoma cells. Molecules 2017;22:2119.
- 97. Babaamer ZY, Sakhri L, Al-Jaber HI, Al-Qudah MA, Abu Zarga MH. Two new taraxasterol-type triterpenes from *Pergularia tomentosa* growing wild in Algeria.

- J Asian Nat Prod Res 2012;14:1137-43.
- 98. Liu JC, Yu LL, Tang MX, Lu XJ, Zhao D, Wang HF, et al. Two new steroidal saponins from the roots of *Cynanchum limprichti*i. J Asian Nat Prod Res 2018;20:875-82.
- Lin M, Han P, Li Y, Wang W, Lai D, Zhou L. Quinoa secondary metabolites and their biological activities or functions. Molecules 2019;24:2512.
- 100. Shi C, Xu MJ, Bayer M, Deng ZW, Kubbutat MH, Wätjen W, et al. Phenolic compounds and their antioxidative properties and protein kinase inhibition from the Chinese mangrove plant *Laguncularia racemosa*. Phytochemistry 2010;71:435-42.
- 101. Bastos KX, Dias CN, Nascimento YM, da Silva MS, Langassner SM, Wessjohann LA, et al. Identification of phenolic compounds from *Hancornia speciosa* (*Apocynaceae*) leaves by UHPLC Orbitrap-HRMS. Molecules 2017;22:143.
- 102. Sánchez-Gutiérrez JA, Moreno-Lorenzana D, Álvarez-Bernal D, Rodríguez-Campos J, Medina-Medrano JR. Phenolic profile, antioxidant and anti-proliferative activities of methanolic extracts from *Asclepias linaria* Cav. Leaves. Molecules 2019;25:54.
- 103. Uchikura T, Tanaka H, Sugiwaki H, Yoshimura M, Sato-Masumoto N, Tsujimoto T, *et al.* Preliminary quality evaluation and characterization of phenolic constituents in *Cynanchi wilfordii* radix. Molecules 2018;23:656.
- 104. Parhira S, Yang ZF, Zhu GY, Chen QL, Zhou BX, Wang YT, *et al. In vitro* anti-influenza virus activities of a new lignan glycoside from the latex of *Calotropis gigantea*. PLoS One 2014;9:e104544.
- 105. Şöhretoğlu D, Masullo M, Piacente S, Kirmizibekmez H. Iridoids, monoterpenoid glucoindole alkaloids and flavonoids from *Vinca major*. Biochem Syst Ecol 2013;49:69-72.
- 106. Ghosh A, Chowdhury N, Chandra G. Plant extracts as potential mosquito larvicides. Indian J Med Res 2012;135:581-98.
- 107. Konno K, Ono H, Nakamura M, Tateishi K, Hirayama C, Tamura Y, *et al.* Mulberry latex rich in antidiabetic sugar-mimic alkaloids forces dieting on caterpillars. Proc Natl Acad Sci U S A 2006;103:1337-41.
- 108. Tan WH, Tao L, Hoang KM, Hunter MD, de Roode JC. The effects of milkweed induced defense on parasite resistance in monarch butterflies, *Danaus plexippus*. J Chem Ecol 2018;44:1040-4.
- 109. Govindarajan M, Sivakumar R, Rajeswary M, Yogalakshmi K. Chemical composition and larvicidal activity of essential oil from *Ocimum basilicum* (L.) against *Culex tritaeniorhynchus*, *Aedes albopictus* and *Anopheles subpictus* (Diptera: Culicidae). Exp Parasitol 2013;134:7-11.
- 110. Hoekou YP, Tchacondo T, Karou SD, Koudouvo K, Atakpama W, Pissang P, *et al.* Ethnobotanical study of latex plants in the maritime region of togo. Pharmacogn

- Res 2016;8:128-34.
- 111. Guo J, Zhou LY, He HP, Leng Y, Yang Z, Hao XJ. Inhibition of 11b-HSD1 by tetracyclic triterpenoids from *Euphorbia kansui*. Molecules 2012;17:11826-38.
- 112. Batool M, Hussain D, Akrem A, Najam-Ul-Haq M, Saeed S, Zaka SM, *et al.* Graphene quantum dots as cysteine protease nanocarriers against stored grain insect pests. Sci Rep 2020;10:3444.
- 113. Harrison RL, Bonning BC. Proteases as insecticidal agents. Toxins (Basel) 2010;2:935-53.
- 114. Asmerom D, Hailu GS, Yimer EM, Bitew H, Kahsay G. Antimicrobial evaluation of latex and TLC fractions from the leaves of *Aloe adigratana* reynolds. Evid Based Complement Alternat Med 2020;2020:8312471.
- 115. Nikkon F, Habib MR, Saud ZA, Karim MR. *Tagetes erecta* Linn. and its mosquitocidal potency against *Culex quinquefasciatus*. Asian Pac J Trop Biomed 2011;1:186-8.
- 116. Dussourd DE. *Theroa zethus* caterpillars use acid secretion of anti-predator gland to deactivate plant defense. PLoS One 2015;10:e0141924.
- 117. Chandrasekaran R, Seetharaman P, Krishnan M, Gnanasekar S, Sivaperumal S. *Carica papaya* (Papaya) latex: A new paradigm to combat against dengue and filariasis vectors *Aedes aegypti* and *Culex quinquefasciatus* (Diptera: Culicidae). 3 Biotech 2018;8:83.
- 118. Kitajima S, Kamei K, Taketani S, Yamaguchi M, Kawai F, Komatsu A, *et al.* Two chitinase-like proteins abundantly accumulated in latex of mulberry show insecticidal activity. BMC Biochem 2010;11:6.
- 119. Parachnowitsch AL, Caruso CM, Campbell SA, Kessler A. *Lobelia siphilitica* plants that escape herbivory in time also have reduced latex production. PLoS One 2012;7:e37745.
- 120. Fürstenberg-Hägg J, Zagrobelny M, Bak S. Plant defense against insect herbivores. Int J Mol Sci 2013;14:10242-97.
- 121. Murthy HN, Joseph KS, Payamalle S, Dalawai D, Ganapumane V. Chemical composition, larvicidal and antioxidant activities of latex from *Garcinia morella* (Gaertn.) Desr. J Parasit Dis 2017;41:666-70.
- 122. Gaupels F, Sarioglu H, Beckmann M, Hause B, Spannagl M, Draper J, *et al.* Deciphering systemic wound responses of the pumpkin extrafascicular phloem by metabolomics and stable isotope-coded protein labeling. Plant Physiol 2012;160:2285-99.
- 123. Megeressa M, Bisrat D, Mazumder A, Asres K. Structural elucidation of some antimicrobial constituents from the leaf latex of *Aloe trigonantha* L.C. Leach. BMC Complement Altern Med 2015;15:270.
- 124. Upadhyay RK. Plant latex: Its toxicity and defense against herbivorous insects: A review. Int J Curr Res 2012;4:5-10.

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